Miltenyi Biotec

CD25 MicroBeads
non-human primate

Order no. 130-091-095

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1. Description

Components	2 mL CD25 MicroBeads, non-human primate: MicroBeads conjugated to monoclonal anti-human CD25 antibodies (isotype: mouse IgG1).
Size	For 10^9 total cells, up to 100 separations.
Product format	CD25 MicroBeads are supplied in buffer containing stabilizer and 0.05 % sodium azide.
Storage	Store protected from light at 2–8 °C. Do not freeze. The expiration date is indicated on the vial label.

This product is applicable for the separation of cells from rhesus monkey (*Macaca mulatta*). The CD25 antibody has been tested to cross-react with cynomolgus monkey (*Macaca fascicularis*). Cross-reactivity has not been tested with other non-human primates.

1.1 Principle of MACS® Separation

First, the CD25$^+$ cells are magnetically labeled with CD25 MicroBeads. Then the cell suspension is loaded onto a MACS® Column which is placed in the magnetic field of a MACS Separator. The magnetically labeled CD25$^+$ cells are retained on the column. The unlabeled cells run through, this cell fraction is depleted of CD25$^+$ cells. After removal of the column from the magnetic field, the magnetically retained CD25$^+$ cells can be eluted as the positively selected cell fraction.

1.2 Background and product applications

CD25 MicroBeads are developed for the separation of non-human primate cells based on the expression of the CD25 antigen. CD25 is expressed on a subset of rhesus monkey peripheral blood mononuclear cells (PBMCs). The CD25 MicroBeads recognize epitope A of the CD25 molecule.

Example application

- Positive selection or depletion of CD25$^+$ cells from rhesus monkey PBMCs, lymphoid tissue or in vitro stimulated lymphocytes.

1.3 Reagent and instrument requirements

- Buffer: Prepare a solution containing phosphate-buffered saline (PBS) pH 7.2, 0.5% bovine serum albumin (BSA), and 2 mM EDTA by diluting MACS BSA Stock Solution (# 130-091-376) 1:20 with autoMACS™ Rinsing Solution (# 130-091-222). Keep buffer cold (4–8 °C). Degas buffer before use, as air bubbles could block the column.

  ▲ **Note:** EDTA can be replaced by other supplements such as anticoagulant citrate dextrose formula-A (ACD-A) or citrate phosphate dextrose (CPD). BSA can be replaced by other proteins such as human serum albumin, human serum, or fetal calf serum. Buffers or media containing Ca2+ or Mg2+ are not recommended for use.

- MACS Columns and MACS Separators: CD25$^+$ cells can be enriched by using MS, LS or XS Columns (positive selection). CD25 MicroBeads can be used for depletion of CD25$^+$ cells on LD, CS or D Columns. Positive selection or depletion can also be performed by using the autoMACS Separator.

Column	Max. number of labeled cells	Max. number of total cells	Separator
Positive selection			
MS	10^7	2×10^8	MiniMACS, OctoMACS, VarioMACS, SuperMACS
LS	10^8	2×10^9	MidiMACS, QuadroMACS, VarioMACS, SuperMACS
XS	10^9	2×10^{10}	SuperMACS
Depletion			
LD	10^8	5×10^8	MidiMACS, QuadroMACS, VarioMACS, SuperMACS
CS	2×10^8		VarioMACS, SuperMACS
D	10^9		SuperMACS
Positive selection or depletion			
autoMACS	2×10^8	4×10^9	autoMACS

▲ **Note:** Column adapters are required to insert certain columns into VarioMACS™ or SuperMACS™ Separator. For details see the respective MACS Separator data sheet.

- (Optional) Fluorochrome-conjugated CD25 antibody recognizing epitope B for flow-cytometric analysis, for example, CD25-PE or CD25-APC.
- (Optional) Propidium iodide (PI) or 7-AAD for the flow-cytometric exclusion of dead cells.
- (Optional) Pre-Separation Filters (# 130-041-407) to remove cell clumps.

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2. Protocol

2.1 Sample preparation

When working with rhesus monkey anticoagulated peripheral blood or buffy coat, peripheral blood mononuclear cells (PBMCs) should be isolated by density gradient centrifugation, for example, using Ficoll-Paque™. For details see section General Protocols in the user manuals or visit www.miltenyibiotec.com/protocols.

▲ **Note:** To remove platelets after density gradient separation, resuspend cell pellet in buffer and centrifuge at 200×g for 10–15 minutes at 20 °C. Carefully aspirate supernatant. Repeat washing step.

When working with tissues, prepare a single-cell suspension by a standard preparation method. For details see section General Protocols in the user manuals or visit www.miltenyibiotec.com/protocols.

▲ **Note:** Dead cells may bind non-specifically to MACS® MicroBeads. To remove dead cells, we recommend using density gradient centrifugation or the Dead Cell Removal Kit (# 130-090-101).



2.2 Magnetic labeling

▲ Work fast, keep the cells cold, and use pre-cooled solutions. This will prevent capping of antibodies on the cell surface and non-specific cell labeling.

▲ Volumes for magnetic labeling given below are for up to 10^7 total cells. When working with fewer than 10^7 cells, use the same volumes as indicated. When working with higher cell numbers, scale up all reagent volumes and total volumes accordingly (e.g. for 2×10^7 total cells, use twice the volume of all indicated reagent volumes and total volumes).

▲ For optimal performance it is important to obtain a single-cell suspension before magnetic separation. Pass cells through 30 µm nylon mesh (Pre-Separation Filters, # 130-041-407) to remove cell clumps which may clog the column.

1. Determine cell number.
2. Wash cells in 1–2 mL of buffer per 10^7 total cells. Centrifuge at 300×g for 10 minutes. Aspirate supernatant completely.
3. Resuspend cell pellet in 80 µL of buffer per 10^7 total cells.
4. Add 20 µL of CD25 MicroBeads per 10^7 total cells.
5. Mix well and refrigerate for 15 minutes at 4–8 °C.

   ▲ **Note:** Working on ice may require increased incubation times. Higher temperatures and/or longer incubation times lead to non-specific cell labeling.
6. (Optional) Add staining antibodies, for example, add 10 µL of CD25-PE per 10^7 total cells, and refrigerate for 5 minutes at 4–8 °C.
7. Wash cells by adding 1–2 mL of buffer per 10^7 cells and centrifuge at 300×g for 10 minutes. Aspirate supernatant completely.
8. Resuspend up to 10^8 cells in 500 µL of buffer.

   ▲ **Note:** For higher cell numbers, scale up buffer volume accordingly.

   ▲ **Note:** For depletion with LD Columns, resuspend up to 1.25×10^8 cells in 500 µL of buffer.
9. Proceed to magnetic separation (2.3).



2.3 Magnetic separation

▲ Choose an appropriate MACS Column and MACS Separator according to the number of total cells and the number of $CD25^+$ cells. For details see table in section 1.3.

Magnetic separation with MS or LS Columns

1. Place column in the magnetic field of a suitable MACS Separator. For details see respective MACS Column data sheet.
2. Prepare column by rinsing with appropriate amount of buffer:

   MS: 500 µL LS: 3 mL
3. Apply cell suspension onto the column.
4. Collect unlabeled cells that pass through the column and wash with appropriate amount of buffer. Perform washing steps by adding buffer three times. Only add new buffer when the column reservoir is empty.

   MS: 3×500 µL LS: 3×3 mL

   Collect total effluent. This is the unlabeled cell fraction.
5. Remove column from the separator and place it on a suitable collection tube.
6. Pipette an appropriate amount of buffer onto the column. Immediately flush out the magnetically labeled cells by firmly pushing the plunger into the column.

   MS: 1 mL LS: 5 mL

   ▲ **Note:** To increase the purity of the magnetically labeled fraction pass the cells over a new, freshly prepared column.

Magnetic separation with XS Columns

For instructions on the column assembly and the separation refer to the XS Column data sheet.

Depletion with LD Columns

1. Place LD Column in the magnetic field of a suitable MACS Separator. For details see LD Column data sheet.
2. Prepare column by rinsing with 2 mL of buffer.
3. Apply cell suspension onto the column.
4. Collect unlabeled cells which pass through and wash with 2×1 mL of buffer. Collect total effluent. This is the unlabeled cell fraction.

Depletion with CS Columns

1. Assemble CS Column and place it in the magnetic field of a suitable MACS Separator. For details see CS Column data sheet.
2. Prepare column by filling and rinsing with 60 mL of buffer. Attach a 22G flow resistor to the 3-way-stopcock of the assembled column. For details see CS Column data sheet.
3. Apply cell suspension onto the column.
4. Collect unlabeled cells which pass through and wash with 30 mL buffer from top. Collect total effluent. This is the unlabeled cell fraction.

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Depletion with D Columns

For instructions on column assembly and separation, refer to the D Column data sheet.

Magnetic separation with autoMACS™ Separator

▲ Refer to the autoMACS™ user manual for instructions on how to use the autoMACS Separator.

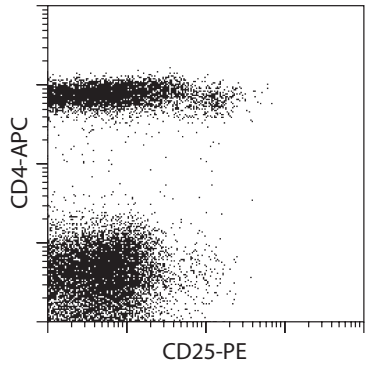
1. Prepare and prime autoMACS Separator.
2. Place tube containing magnetically labeled cells in the autoMACS Separator. Choose a separation program according to the recommendations in the autoMACS user manual.

   ▲ **Note:** Program choice depends on the isolation strategy, the strength of magnetic labeling and the frequency of magnetically labeled cells. For details see autoMACS user manual, section autoMACS Cell Separation Programs.

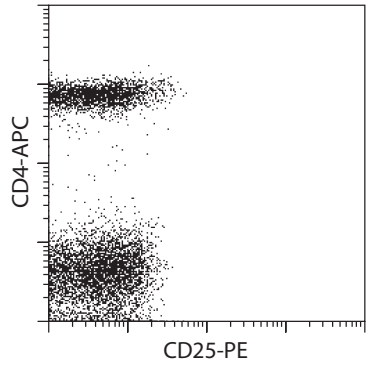
3. Example of a separation using CD25 MicroBeads

$CD25^+$ cells were depleted from rhesus monkey PBMCs using CD25 MicroBeads, a MidiMACS™ Separator and an LD Column. Cells were stained with CD25-PE and CD4-APC. Cell debris and dead cells were excluded from the analysis based on scatter signals and PI fluorescence.

PBMCs before separation



$CD25^-$ cell fraction



Refer to **www.miltenyibiotec.com** for all data sheets and protocols. Miltenyi Biotec provides technical support worldwide. Visit www.miltenyibiotec.com for local Miltenyi Biotec Technical Support contact information.

Warnings

Reagents contain sodium azide. Under acidic conditions sodium azide yields hydrazoic acid, which is extremely toxic. Azide compounds should be diluted with running water before discarding. These precautions are recommended to avoid deposits in plumbing where explosive conditions may develop.

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