

CliniMACS Prodigy® Adherent Cell Culture

Differentiation of human pluripotent stem cells into cardiomyocytes

Application

The CliniMACS Prodigy Adherent Cell Culture process enables expansion and differentiation of human pluripotent stem cells (PSCs) in a closed and automated system. In combination with the ready-to-use, xeno-free StemMACS™ CardioDiff Kit XF, human, it supports robust and scalable differentiation of human PSCs into cardiomyocytes in just 8 days. This application sheet provides an entire process overview in addition to highlighting information about the required materials. In addition, it illustrates the setup of the tubing set CliniMACS Prodigy TS 730 and the key performance data.

Specifications

Process capacity:	scalable
Number of PSCs for differentiation:	approx. 1.6×10 ⁸ cells*
Number of final cardiomyocytes:	up to 2.9×10 ⁸ cells
Total process time:	8 days
Total hands-on time:	approx. 10 h

* The optimal initial seeding density for differentiation is cell line dependent. This is the cell number we used for our cell line in a CellSTACK 1 Chamber.

Products

Consumables	Amount required
CliniMACS Prodigy Instrument	1 piece
CliniMACS Prodigy TS 730	1 set
StemMACS CardioDiff Kit XF, human	4 kits
PBS	3 L
1 m Tube Extension	1 piece
MPC male Adapter 1/4" ID	1 piece
In-Line Filter Adapter (200 μM)	1 piece

Additional materials	Amount required
Corning® 33 mm Polyethylene Filling Cap with a Female MPC Polycarbonate with a 1/4 (6.4 mm) ID Coupling and a Male MPC Polycarbonate End Cap	1 piece
Corning CellSTACK [®] – 1 Chamber	1 piece
Corning 1000 mL Easy Grip Polystyrene Storage Bottles with Dip Tube, with 0.2 µm MLL/FLL Filter*	2 pieces
Flexboy Bag 500 mL, Inlet: Luer Lock male + cap, Outlet: Luer Lock female + cap, Sartorius	5 pieces
Gibco™ TrypLE™ Select Enzyme (10×), no phenol red, Thermo Fisher	100 mL
Gibco Soybean Trypsin Inhibitor, powder, Thermo Fisher	60 mL (5 mg/mL)
Biolaminin 521 LN (LN521), 500 μg, BioLamina	2 vials

* Used as alternative vessels for StemMACS CardioDiff Kit XF, human

Process overview for cardiomyocyte differentiation

	Tubing set installation and priming
Pre-process	▼
(day 0)	Tubing set blocking with culture medium
	▼
	Coating of one CellSTACK – 1 Chamber with LN521
Coating and inoculation	\mathbf{v}
(uay v)	Inoculation of PSCs in one CellSTACK – 1 Chamber in Mesoderm Induction Medium (MIM)
	Day 1 – Rinsing cells with PBS and medium change with
	Cardiac Cultivation Medium (CCM)
Cultivation and medium change	▼
(day 1–3)	Day 2 – Rinsing cells with PBS and medium change with
	Dav 3 – Rinsing cells with PBS and medium change with CCM
	▼
Cultivation and medium change	Daily medium change with CCM
(day 4–7)	▼
	Semi-automated harvest and filtration of cardiomyocytes
	Semi-automated harvest and filtration of cardiomyocytes
Harvest and final formulation (day 8)	Semi-automated harvest and filtration of cardiomyocytes Sample collection for QC and cell counting
Harvest and final formulation (day 8)	Semi-automated harvest and filtration of cardiomyocytes Sample collection for QC and cell counting Storage of cells in target cell bag
Harvest and final formulation (day 8)	Semi-automated harvest and filtration of cardiomyocytes Sample collection for QC and cell counting Storage of cells in target cell bag
Harvest and final formulation (day 8) Post-process	Semi-automated harvest and filtration of cardiomyocytes Sample collection for QC and cell counting Storage of cells in target cell bag Tubing set deinstallation
Harvest and final formulation (day 8) Post-process (day 8)	Semi-automated harvest and filtration of cardiomyocytes Sample collection for QC and cell counting Storage of cells in target cell bag Tubing set deinstallation
Harvest and final formulation (day 8) Post-process (day 8) Quality control	Semi-automated harvest and filtration of cardiomyocytes Sample collection for QC and cell counting Storage of cells in target cell bag Tubing set deinstallation Cardiomyocyte characterization
Harvest and final formulation (day 8) Post-process (day 8) Quality control (day 8)	Semi-automated harvest and filtration of cardiomyocytes Sample collection for QC and cell counting Storage of cells in target cell bag Tubing set deinstallation Cardiomyocyte characterization
Harvest and final formulation (day 8) Post-process (day 8) Quality control (day 8) Continuous cultivation (> day 8)	Semi-automated harvest and filtration of cardiomyocytes Sample collection for QC and cell counting Storage of cells in target cell bag Tubing set deinstallation Cardiomyocyte characterization Cardiomyocyte characterization

8 days for total process



Take a look at our protocol for further information on immunophenotyping using flow cytometry

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Principle of the cardiomyocyte differentiation process using the CliniMACS Prodigy



CliniMACS Prodigy TS 730 setup for cardiomyocyte differentiation



Performance data



Approximately 1.6×10⁸ human PSCs were used for differentiation into cardiomyocytes using the CliniMACS Prodigy Adherent Cell Culture process with the StemMACS CardioDiff Kit XF, human. The differentiation took place in a CellSTACK – 1 Chamber from day 0 to day 8 with varying medium. (day 0 - Mesoderm Induction Medium, day 1- Cardiac Cultivation Medium, day 2- Cardiac Induction Medium, days 3-8 Cardiac Cultivation Medium). After 8 days of differentiation, the expression levels of cardiomyocyte markers α-Actinin (ACTN) (A) and Cardiac Troponin T (cTNT) (B) in the resulting cells were analyzed by flow cytometry, revealing a differentiation efficiency of approx. 78%. Furthermore, up to 2.9×10⁸ ACTN⁺ cardiomyocytes could be harvested from a CellSTACK - 1 Chamber and larger external culture vessels can be employed for a scalable cardiomyocyte differentiation processes.



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