

Large-scale manufacturing of transduced T cells

CliniMACS Prodigy® T Cell Transduction – Large Scale

Application

Fully automated labeling, enrichment, activation, transduction, and expansion of human T cells from frozen or fresh leukapheresis for the production of gene-engineered T cells.

This application sheet gives an overview of the specifications and materials required to perform the T Cell Transduction – Large Scale (TCT–LS) Process. Furthermore, it provides an overview of the setup for the tubing set, general workflow, and internal performance data.

Specifications

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Process name:	T Cell Transduction – Large Scale
Selection capacity:	Up to 3×10° labeled cells within 10×10° WBCs
Sample volume for selection:	Fresh: 50–600 mL Frozen: 50–300 mL
TransAct™ Large Scale stimulation capacity:	4×10 ⁸ T cells recommended
Expansion capacity:	3×10 ⁷ T cells/mL in max. 600 mL culture volume
Final product harvest volume:	Four different harvest types available:
	 Optional rebuffering step
	Optional transfer of cells into Target Cell Bag
Process time:	12 days recommended

Products required*

CliniMACS® and MACS® GMP Products	Amount required		
CliniMACS Prodigy with T Cell Transduction – Large Scale application software	1		
CliniMACS CD4 and CD8 Reagent/GMP MicroBeads	1 piece each		
CliniMACS CD62L Reagent/GMP MicroBeads	1 piece		
CliniMACS Prodigy TS 620	1 piece		
CliniMACS PBS/EDTA Buffer (3 L)	1 piece		
TexMACS™ GMP Medium	5×2 L bags		
MACS GMP Recombinant Human IL-7 and IL-15	1 vial each per one 2 L medium bag		
MACS GMP Recombinant Human IL-2	<1 vial per one 2 L medium bag		
MACS GMP T Cell TransAct Large Scale	1 vial		

Additional material / equipment*
Viral vector
3-way Tube Adapter (Miltenyi Biotec)
Triple Sampling Adapter (Miltenyi Biotec)
Sterile water, syringes, hypodermic needles
MACS GMP Vectofusin®-1
Formulation solution (e.g. CliniMACS Formulation Solution)
Human serum albumin
Human AB serum
Sterile tube welder
Uninterruptable power supply
CO ₂ and compressed air supply
Cell counter and/or flow cytometer
*Depending on the charge protocol same consumables might not be

^{*}Depending on the chosen protocol some consumables might not be needed. Contact your Miltenyi Biotec representative for support.

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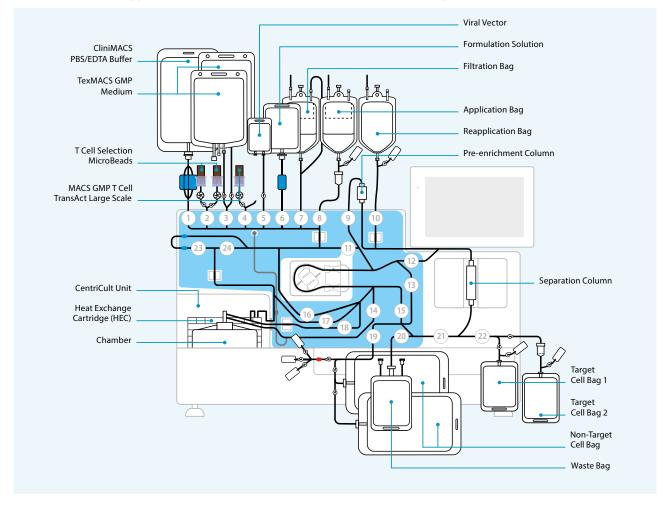
Process overview

Pre-process	Tubing set installation and priming Connection of fresh or frozen cell product			
T cell enrichment	T cell enrichment (CD4/CD8)			
Activation	Stimulation of T cells with MACS GMP T Cell TransAct Large Scale			
Viral transduction	Lentiviral or retroviral (+Vectofusin-1)			
Cell expansion	Expansion in TexMACS Medium with AB serum, IL-7, and IL-15			
Cell harvest and formulation	Four different harvest types available offering optional rebuffering step and optional transfer of cells into Target Cell Bag			
Post-process	Tubing set deinstallation Tubing set deinstallation			

12 days recommended**

 $[\]hbox{** The duration of the T Cell Transduction - Large Scale Process depends on the required cell numbers.}$

CliniMACS Prodigy TS 620 setup for T Cell Transduction – Large Scale



Performance data

	Start product	Isolated cells		Final product						
Conditions	CD4 ⁺ and CD8 ⁺ T cells (%)	CD4 ⁺ and CD8 ⁺ T cells (%)	CD3 ⁺ cells viability (%)	CD4 ⁺ and CD8 ⁺ T cells (%)	CD3 ⁺ T cells (×10 ¹⁰)	CAR ⁺ T cells (%)	Viability (%)	CAR ⁺ T cell number (×10 ⁹)		
Untransduced										
Fresh LP, IL-7 and IL-15 (n = 1 separation, n = 2 culture)	57.79	91.00	98.81	96.69±0.08	1.80±0.11	-	95.12±0.39	-		
Transduced										
Fresh LP, IL-7 and IL-15 (n = 2)	44.61 ±6.57	86.60 ±3.81	98.38 ±0.56	97.31 ±0.96	1.71 ±0.05	29.55 ±0.79	96.63 ±0.08	5.03 ±0.27		
Frozen LP, IL-7 and IL-15 (n = 3)	*48.30 ±4.02	84,76 ±2.25	94,43 ±3.06	96.40 ±1.25	1.93 ±0.15	41.98 ±13.70	94.79 ±1.94	7.67 ±1.87		
Fresh LP, IL-2 (n = 1)	36.91	74.92	98.06	90.63	1.62	29.69	91.30	4.81		
Frozen LP, IL-2 (n = 1)	*53.98	81.82	90.54	97.11	1.46	58.06	98.08	8.48		

^{*}Sample taken after thawing from application bag

Table 1: Internal data showing performance data of several TCT-LS runs. Fresh or frozen leukapheresis from healthy donors was used as a starting product. The culture was performed with media supplementation of IL-2, or a combination of IL-7 and IL-15. Two runs were performed without transduction, and the remaining seven with lentiviral transduction. Following T cell isolation, high T cell purities and viabilities were observed in all conditions. High expansion rates and viabilities were also observed after culture. The different transduction efficiencies between process runs are most likely a donor variability effect. This data demonstrates that the TCT-LS is robust enough to deal with variable conditions and still produce a good result.

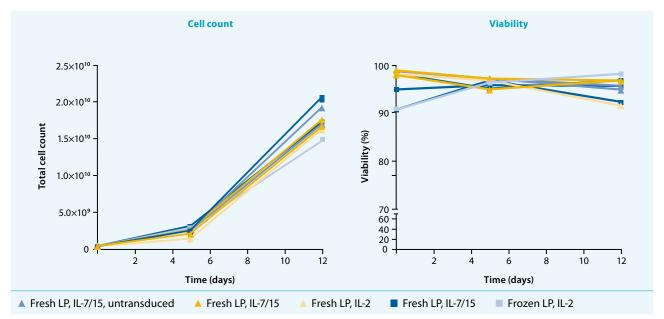


Figure 1: Internal data showing the cell expansion and viability during a complete TCT-LS Process. Different starting products and culture conditions were used, and remarkedly, no huge difference between the variable conditions were observed, demonstrating the robustness of the process. As depicted in the left graph, cell expansion markedly increased after day six, reaching up to 2.05×10^{10} T cells in the end product, whilst viability remained >90% during the whole process.



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In the EU, the CliniMACS System components are available as CE-marked medical devices for their respective intended use, unless otherwise stated. The CliniMACS Reagents and Biotin Conjugates are intended for *in vitro* use only and are not designated for therapeutic use or direct infusion into patients. The CliniMACS Reagents in combination with the CliniMACS System are intended to separate human cells. Miltenyi Biotec as the manufacturer of the CliniMACS System does not give any recommendations regarding the use of separated cells for therapeutic purposes and does not make any claims regarding a clinical benefit. For the manufacturing and use of target cells in humans, the national legislation and regulations – e.g. for the EU the Directive 2004/23/EC ("human tissues and cells"), or the Directive 2002/98/EC ("human blood and blood components") – must be followed. Thus, any clinical application of the target cells is exclusively within the responsibility of the user of a CliniMACS System.

In the US, the CliniMACS CD34 Reagent System, including the CliniMACS Plus Instrument, CliniMACS CD34 Reagent, CliniMACS Tubing Sets TS and LS, and the CliniMACS PBS/EDTA Buffer, is FDA approved as a Humanitarian Use Device (HUD), authorized by U.S. Federal law for use in the treatment of patients with acute myeloid leukemia (AML) in first complete remission. The effectiveness of the device for this indication has not been demonstrated. Other products of the CliniMACS Product Line are available for use only under an approved Investigational New Drug (IND) application, Investigational Device Exemption (IDE) or FDA approval. CliniMACS GMP MicroBeads are for research use and ex vivo cell processing only.

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