

FFPE Tissue Dissociation Kit for RNA Profiling

Order no. 130-134-089

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1. Description

This product is for research use only.

Components	1 vial of Enzyme D (lyophilized powder) 2.5 mL Enzyme P	
	13 mL Buffer S (20×)	
Size	For 25 digestions. The specified number of digestions is valid when digesting up to two 25 μ m tissue scrolls in 2.5 mL enzyme mix following the protocol in chapter 2.2.	
Storage	Upon arrival store Enzyme P aliquoted at -20 °C. Store all other components at +2 to +8 °C. Reconstitute all components before the date	

Store all other components at +2 to +8 °C. Reconstitute all components before the date indicated on the box label. For information about reconstitution and storage after reconstitution of the lyophilized components refer to chapter 2.1.

1.1 Principle of the FFPE Tissue Dissociation Kit for RNA Profiling

Formalin-fixed paraffin-embedded (FFPE) tissue sections from various tissues and species, for example, human tumor, skin, or lymph node samples, are dissociated into single-nucleus suspensions by combining mechanical dissociation with enzymatic degradation of the extracellular matrix. This maintains the structural integrity of tissues. The FFPE tissue sections are deparaffinized and rehydrated. Then, samples are dissociated using the kit components and the gentleMACS[™] Octo Dissociator with Heaters. After dissociation, the sample is applied to a filter to remove any remaining larger particles from the single-nucleus suspension.

1.2 Background information

The FFPE Tissue Dissociation Kit for RNA Profiling has been optimized for the gentle, rapid, and effective generation of singlenucleus suspensions from FFPE tissue samples. Thus, a high yield of single-nuclei can be obtained. Extracted nuclei can be subsequently counted by flow cytometry or microscopy and be used for molecular applications like single-nucleus RNA sequencing.

1.3 Applications

- Dissociation of FFPE tissue sections from various tissues into single-nucleus suspensions for subsequent single-nucleus RNA sequencing
- Enumeration of nuclei by flow cytometry or fluorescence microscopy

1.4 Reagent and instrument requirements

- gentleMACS Octo Dissociator with Heaters (# 130-096-427)
- gentleMACS C Tubes (25 tubes (# 130-093-237))
- Pre-Separation Filters (30 μm) (# 130-041-407)
- Centrifuge (cooled to +4 °C)
- Glass Pasteur pipette
- Xylol or xylol substitute
- 50%, 70%, and 100% ethanol
- Sterile, RNase-free distilled water
- PBS
- 1 M RNase-free Tris, pH 8
- 10% RNase-free BSA Stock Solution
- RNase inhibitor (40 U/μL)

2. Protocol

▲ For details on the use of the gentleMACS Octo Dissociator with Heaters, refer to the gentleMACS Dissociators user manuals.

 \blacktriangle Volumes given below are for up to two FFPE tissue scrolls (25 μm each) in 2.5 mL enzyme mix per gentleMACS C Tube.

▲ Separation of tumor cells from healthy cells, e.g., with Anti-Cytokeratin MicroBeads, human (# 130-123-094), requires a specific antigen retrieval step (incubation at +80 °C) and is not part of this protocol. For this application, use the FFPE Tissue Dissociation Kit (# 130-188-052) and refer to the respective protocol.

2.1 Reagent preparation

▲ Handle all buffers and reconstituted enzymes under sterile conditions to avoid contamination.

▲ Do not prepare a stock solution containing both Enzyme D and Enzyme P.

- Prepare Buffer S (1×) by mixing, for example, 500 µL Buffer S (20×) with 9.5 mL sterile distilled water under sterile conditions. For long-term storage, store Buffer S (1×) at +2 to +8 °C.
- 2. Prepare Enzyme D by reconstitution of the lyophilized powder in the vial with 3 mL of Buffer S (1×). Invert vial after closing and wait 5–10 minutes while inverting every minute to

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dissolve the pellet. Do not resuspend by pipetting or vortexing. Prepare aliquots of appropriate volume to avoid repeated freeze-thaw-cycles. Store aliquots at -20 °C. This solution is stable for 6 months after reconstitution.

- 3. Prepare aliquots of Enzyme P of appropriate volume to avoid repeated freeze-thaw-cycles. Store aliquots at -20 °C. This solution is stable for 6 months.
- 4. Prepare resuspension buffer for dissociated FFPE samples $(550 \ \mu L \ per \ dissociation \ run)$, e.g., $0.5 \times \ PBS$ containing 50 mM RNase-free Tris, pH 8, 0.02% RNase-free BSA, and $0.2 \ U/\mu L$ RNase inhibitor. Table 1 shows the required volumes of the respective stock solutions for one or four preparations. An overfill of 10% is included. Adapt the calculations for the target concentrations if working with other stock solutions.

▲ Note: This kit has been validated using the indicated resuspension buffer. Use of any other resuspension buffer is at own risk.

	For one dissociation run	For four dissociation runs
1× PBS	275 μL	1100 μL
1 M RNase-free Tris, pH 8	28 μL	112 μL
10% RNase-free BSA	1 μL	4 μL
RNase inhibitor (40 U/µL)	3 µL	12 μL
Sterile, RNase-free water	243 μL	972 μL

 Table 1: Preparation of resuspension buffer for one dissociation run or four dissociation runs.

2.2 Dissociation of FFPE tissue

▲ The volumes listed below are based on one dissocation run of up to two FFPE tissue scrolls. Increase the volumes based on the number of dissociation runs. Use a maximum of two FFPE tissue scolls per C Tube.

 \blacktriangle Cool down centrifuge to +4 °C.

▲ Prewarm an aliquot of 2.35 mL Buffer S (1×) for at least 10 min at +37 °C before use for step 9.

▲ Place an aliquot of 1 mL Buffer S (1×) on ice.

▲ This protocol has been validated with xylene. Use xylene substitutes at own risk.

Always use glass Pasteur pipettes as samples may stick to plastic pipettes.

1. Prepare up to two 25 μ m tissue scrolls from an FFPE block, remove excessive paraffin, and transfer the scrolls into a gentleMACS C Tube.

▲ Note: The yield from two 25 µm scrolls may not be sufficient due to differences in cell density and tissue size in the FFPE block. If the final number of nuclei is not sufficient, pooling of multiple runs is needed. It is recommended to perform a test run to determine the number of scrolls required for a specific target nuclei number.

▲ Note: In case a tissue section is already mounted on a slide, place the slide including the section in an appropriate container for deparaffination and rehydration (steps 2–8). Be aware that much higher volumes are needed to wet samples.

2. Add 3 mL of xylene or xylene substitute to the gentleMACS C Tube, incubate for 10 minutes at room temperature, and aspirate liquid completely using a glass Pasteur pipette.

▲ Note: For optimal aspiration of liquids, move the section(s) carefully with the glass Pasteur pipette tip along the tube wall above the liquid level, then

aspirate the liquid completely without touching or removing the scrolls.

- 3. Repeat step 2 twice for a total of three washes with xylene or xylene substitute.
- 4. Add 3 mL of 100% ethanol to the gentleMACS C Tube, incubate for 30 seconds at room temperature, and aspirate liquid completely using a glass Pasteur pipette.

▲ Note: For optimal aspiration of liquids, move the section(s) carefully with the glass Pasteur pipette tip along the tube wall above the liquid level, then aspirate the liquid completely without touching or removing the scrolls.

5. Add 1 mL of 100% ethanol to the gentleMACS C Tube, incubate for 30 seconds at room temperature, and aspirate liquid completely using a glass Pasteur pipette.

▲ Note: For optimal aspiration of liquids, move the section(s) carefully with the glass Pasteur pipette tip along the tube wall above the liquid level, then aspirate the liquid completely without touching or removing the scrolls.

6. Add 1 mL of 70% ethanol to the gentleMACS C Tube, incubate for 30 seconds at room temperature, and aspirate liquid completely using a glass Pasteur pipette.

▲ Note: For optimal aspiration of liquids, move the section(s) carefully with the glass Pasteur pipette tip along the tube wall above the liquid level, then aspirate the liquid completely without touching or removing the scrolls.

7. Add 1 mL of 50% ethanol to the gentleMACS C Tube, incubate for 30 seconds at room temperature, and aspirate liquid completely using a glass Pasteur pipette.

▲ Note: For optimal aspiration of liquids, move the section(s) carefully with the glass Pasteur pipette tip along the tube wall above the liquid level, then aspirate the liquid completely without touching or removing the scrolls.

- 8 Add 1 mL of deionized water to the gentleMACS C Tube, incubate for 30 seconds at room temperature, and aspirate liquid completely using a glass Pasteur pipette.
- 9. Prepare enzyme mix by mixing 2.35 mL of prewarmed $1 \times$ Buffer S, 100 µL of Enzyme D, and 65 µL of Enzyme P. Transfer dissociation mix into the gentleMACS C Tube and close it tightly.
- 10. Attach the C Tube to the gentleMACS Octo Dissociator with Heaters and place the Heater Unit on the C Tube. Run the gentleMACS program 37C_FFPE_1.
- 11. After termination of the program, detach the C Tube from the gentleMACS Octo Dissociator with Heaters and place it on ice.
- 12. (Optional) Perform a short spin up to $300 \times g$ to collect the sample at the bottom of the tube. Resuspend the cells.
- 13. Apply the cell suspension to a Pre-Separation Filter (30 $\mu m)$ placed on a 15 mL tube on ice.
- 14. Rinse the used C Tube with 1 mL ice-cold 1× Buffer S and apply to the Pre-Separation Filter (30 $\mu m).$
- 15. Discard the filter and centrifuge the nuclei suspension at 850×g at +4 °C for 5 minutes. Aspirate supernatant completely.
- 16. Resuspend the nuclei pellet in 0.5 mL of an appropriate resuspension buffer for downstream application.

▲ Note: A nuclei pellet might not be visible due to the low amount of nuclei.

17. Process nuclei immediately for further applications, e.g., nuclei counting or single-nucleus RNA sequencing.

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