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Warnings

Reagents contain sodium azide. Under acidic conditions sodium azide yields hydrazoic acid, which is extremely toxic. Azide compounds should be diluted with running water before discarding. These precautions are recommended to avoid deposits in plumbing where explosive conditions may develop.

1. Description

This product is for research use only.

Components	1 mL Tissue Stem Cell Pre-Enrichment Cocktail, mouse: MicroBeads conjugated to monoclonal anti-mouse antibodies.
Capacity	For 5×10^8 total cells, up to 50 separations.
Product format	Cocktail is supplied in buffer containing stabilizer and 0.05% sodium azide.
Storage	Store protected from light at 2–8 °C. Do not freeze. The expiration date is indicated on the vial label.

1.1 Principle of the MACS® Separation

First, the committed and differentiated cells are magnetically labeled with a cocktail of monoclonal antibodies conjugated with MACS® MicroBeads. Then, the cell suspension is loaded onto a MACS Column, which is placed in the magnetic field of a MACS Separator. The magnetically labeled cells are retained within the column. The unlabeled tissue stem cells run through. After removing the column from the magnetic field, the magnetically retained lineage cells can be eluted.

1.2 Background information

The Tissue Stem Cell Pre-Enrichment Kit, mouse has been designed for the pre-enrichment of untouched tissue resident cell populations, e.g., stem and progenitor cells, from primary specimens. All primary tissues are vascularized and infiltrated by heterogeneous lymphocyte subpopulations, red blood cells, and endothelial cells. The level of heterogeneity is highly dependent on multiple factors like tissue type, age, and disease state.

The isolation of target cells from primary tissues, such as organ specific stem and progenitor cells, is frequently hampered by the initial low frequency of these cells. This may lead to long isolation times and impaired purities. By using the Tissue Stem Cell Pre-Enrichment Kit, mouse all generic infiltrating cells, such as red and white blood cells as well as endothelial cells are efficiently depleted resulting in a significant pre-enrichment of target cells for downstream analysis or sorting.

1.3 Applications

- Enrichment of untouched tissue–resident stem and progenitor cells.

1.4 Reagent and instrument requirements

- Buffer: Prepare a solution containing phosphate-buffered saline (PBS), pH 7.2, 0.5% bovine serum albumin (BSA), and 2 mM EDTA by diluting MACS® BSA Stock Solution (# 130-091-376) 1:20 with autoMACS® Rinsing Solution (# 130-091-222). Keep buffer cold (2–8 °C). Degas buffer before use, as air bubbles could block the column. Always use freshly prepared buffer.

Do **not use** autoMACS Running Buffer or MACSQuant® Running Buffer as they contain a small amount of sodium azide that could affect the results.

▲ **Note:** EDTA can be replaced by other supplements such as anticoagulant citrate dextrose formula-A (ACD-A) or citrate phosphate dextrose (CPD). BSA can be replaced by other proteins such as mouse serum albumin, mouse serum, or fetal bovine serum (FBS). Buffers or media containing Ca^{2+} or Mg^{2+} are not recommended for use.

- MACS Columns and MACS Separators: For optimal purity and recovery the use of an LS Column is strongly recommended. Depletion can also be performed by using the autoMACS Pro Separator.

Column	Max. number of labeled cells	Max. number of total cells	Separator
LS	4×10^7	5×10^7	MidiMACS, QuadroMACS, VarioMACS, SuperMACS II
autoMACS	5×10^7	10^8	autoMACS Pro

▲ **Note:** Column adapters are required to insert certain columns into the VarioMACS™ or SuperMACS™ II Separators. For details refer to the respective MACS Separator data sheet.

- (Optional) Fluorochrome-conjugated antibodies for flow cytometric analysis, e.g., Anti-Ter-119-APC, mouse (# 130-102-290), CD31-APC, mouse (# 130-102-571), CD45-APC, mouse

(# 130-102-544), Labeling Check Reagent-VioBlue® (# 130-095-087). For more information about antibodies refer to www.miltenyibiotec.com/antibodies.

- gentleMACS™ Dissociator (# 130-093-235), gentleMACS Octo Dissociator (# 130-095-937), or gentleMACS Octo Dissociator with Heaters (# 130-096-427)
- gentleMACS C Tubes (# 130-093-237, # 130-096-334)
- Pre-Separation Filters, 70 μm (# 130-095-823) to remove cell clumps.
- (Optional) Propidium Iodide Solution (# 130-093-233) or 7-AAD for flow cytometric exclusion of dead cells.

2. Protocol

2.1 Sample preparation

For preparation of a single-cell suspension from solid tissues it is recommended to use gentleMACS Dissociators in combination with tissue-specific dissociation kits.

▲ Dead cells may bind non-specifically to MACS MicroBeads. To remove dead cells, we recommend using density gradient centrifugation or the Dead Cell Removal Kit (# 130-090-101).



2.2 Magnetic labeling

▲ Work fast, keep cells cold, and use pre-cooled solutions. This will prevent capping of antibodies on the cell surface and non-specific cell labeling.

▲ Volumes for magnetic labeling given below are for up to 10^7 cells from dissociated tissue. When working with less cells, use the same volumes as indicated. When working with higher amounts, scale up all reagent volumes and total volumes accordingly to the final cell number of dissociated tissue (e.g. for 2×10^7 cells, use twice the volume of all indicated reagent volumes and total volumes). It is strongly recommended to determine the actual cell number from dissociated tissue prior to magnetic labeling.

▲ For optimal performance it is important to obtain a single-cell suspension before magnetic labeling. Pass cells through 70 μm nylon mesh (Pre-Separation Filters, 70 μm, # 130-095-823) to remove cell clumps which may clog the column. Moisten filter with buffer before use.

▲ The recommended incubation temperature is 2–8 °C. Higher temperatures and/or longer incubation times may lead to non-specific cell labeling. Working on ice may require increased incubation times.

1. Determine cell number.
2. Centrifuge cell suspension at $300 \times g$ for 5 minutes. Aspirate supernatant completely.
3. Resuspend cell pellet in 80 μL of buffer per 10^7 total cells.
4. Add 20 μL of Tissue Stem Cell Pre-Enrichment Cocktail per 10^7 total cells.
5. Mix well and incubate for 15 minutes in the refrigerator (2–8 °C).

6. Adjust volume to 500 μL with buffer for up to 5×10^7 total cells.

▲ **Note:** If more tissue was used split sample into multiple tubes.

7. Proceed to magnetic separation (2.3).



2.3 Magnetic separation: Depletion of lineage-positive cells

▲ Choose an LS Column and MACS Separator. For details refer to table in section 1.4.

▲ Always wait until the column reservoir is empty before proceeding to the next step.

Magnetic separation with LS Columns

1. Place LS Column in the magnetic field of a suitable MACS Separator. For details refer to the respective MACS Column data sheet.
2. Prepare column by rinsing with 3 mL of buffer.
3. Apply cell suspension onto the column. Collect flow-through containing unlabeled cells, representing the enriched stem and progenitor cells.
4. Wash column with 2×1 mL of buffer. Collect unlabeled cells that pass through and combine with the flow-through from step 3.

▲ **Note:** Perform washing steps by adding buffer aliquots as soon as the column reservoir is empty.
5. (Optional) Remove column from the separator and place it on a suitable collection tube. Pipette 3 mL of buffer onto the column. Immediately flush out the magnetically labeled non-progenitor cells by firmly pushing the plunger into the column.

Magnetic separation with the autoMACS® Pro Separator

▲ Refer to the respective user manual for instructions on how to use the autoMACS® Pro Separator.

▲ Buffers used for operating the autoMACS Pro Separator should have a temperature of ≥ 10 °C.

1. Prepare and prime the instrument.
2. Apply tube containing the sample and provide tubes for collecting the labeled and unlabeled cell fractions. Place sample tube in row A of the tube rack and the fraction collection tubes in rows B and C.
3. For a standard separation choose one of the following programs:

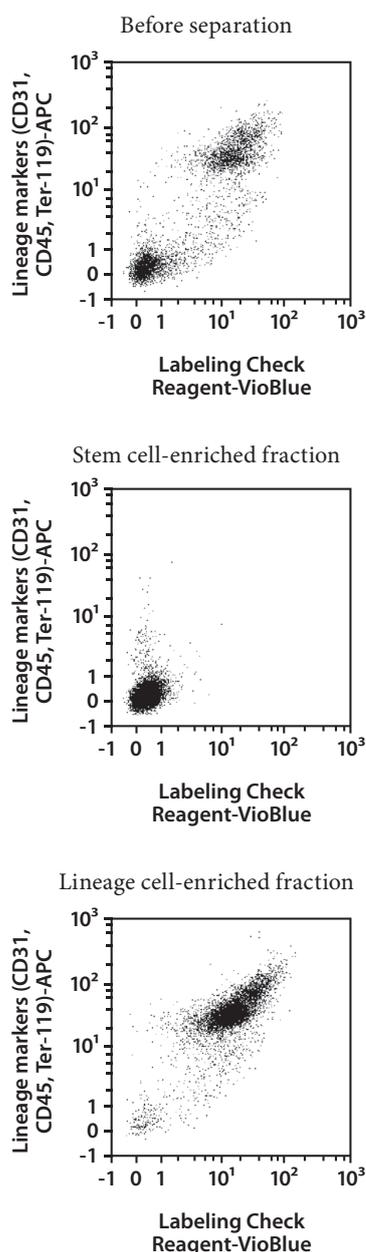
Depletion: Depletes

Collect negative fraction in row B of the tube rack. This fraction represents the enriched stem and progenitor cells.

4. (Optional) Collect positive fraction from row C of the tube rack. This fraction represents the magnetically labeled non-target cells.

3. Example of a separation using the Tissue Stem Cell Pre-Enrichment Kit

Mouse mammary gland was dissociated using the Adipose Tissue Dissociation Kit, mouse and rat (# 130-105-808) in combination with the gentleMACS Octo Dissociator. Subsequently, tissue-resident stem and progenitor cells were pre-enriched using the Tissue Stem Cell Pre-Enrichment Kit, mouse. Cells were fluorescently stained with lineage markers (CD31, CD45, Ter-119-APC conjugated to APC) and Labeling Check Reagent-VioBlue and analyzed by flow cytometry using the MACSQuant® Analyzer. Cell debris and dead cells were excluded from the analysis based on scatter signals and propidium iodide fluorescence.



Refer to www.miltenyibiotec.com for all data sheets and protocols. Miltenyi Biotec provides technical support worldwide. Visit www.miltenyibiotec.com/local to find your nearest Miltenyi Biotec contact.

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