



Miltenyi Biotec



Immunophenotyping

Identification of 8 immune cell subsets from mouse spleen using 14-color panel

Background

Flow cytometry has become the method of choice for immunophenotyping and analysis of specific cellular subsets. Using antibodies against selective markers, it provides a quick overview of cell types that constitute a sample. At the same time it allows a thorough analysis on single-cell level for in-depth characterization. Using multiple markers simultaneously increases the number of parameters that can be analyzed per run and decreases the amount of starting material required to perform an assay. This can be critical for precious sample material and long-term immune monitoring studies. In this application note, we demonstrate a 14-color immunophenotyping of mouse splenocytes allowing for the simultaneous identification and analysis of 8 different immune cell subsets, including viability and activation state of the cells. The data was acquired using the MACSQuant® Analyzer 16, a compact and reliable benchtop flow cytometer equipped with three lasers for high content analysis.

Materials and methods

Splenocytes were labeled with CD11c-VioBlue®, MHCII-VioGreen™, CD11b-BV570™, CD64-BV605™, NK1.1-BV650™, CD172a-Vio® Bright 515, XCR1-PE, CD19-PE-Vio 615, CD3-PerCP, F4/80-PerCP-Vio 700, Siglec-H-PE-Vio 770, CD86-APC, BD Horizon™ Fixable Viability Stain (FVS) 700 and CD45-APC-Vio 770. Data was acquired on the MACSQuant Analyzer 16 using MACSQuantify™ Software. The markers used to identify different immune cell populations are described in table 1.

Cell staining protocol

- Resuspend 2×10^6 splenocytes in 100 μ L of PEB Buffer (phosphate buffered saline, pH 7.2, 2 mM EDTA, 0.5% BSA).
- Centrifuge at 300xg for 10 minutes, aspirate supernatant completely and resuspend cells in 100 μ L of PBS. Repeat washing step once.

- Stain with Fixable Viability Stain (FVS) according to the manufacturer's instructions.
- Resuspend cells in 100 μ L PEB buffer.
- Stain cells with conjugated antibodies according to the manufacturer's instructions.
- Add 1 mL of PEB Buffer and centrifuge at 300xg for 10 min and aspirate supernatant. Repeat washing step once.
- Resuspend pellet in 500 μ L of PEB Buffer.

| Cell type | Function | Phenotype |
|-------------|---|--|
| B cells | Antibody production and adaptive immunity | CD45 ⁺ , F4/80 ⁻ , CD64 ⁻ , CD3 ⁻ , NK1.1 ⁻ , CD19 ⁺ |
| NK cells | Viral and cancer cell clearance | CD45 ⁺ , F4/80 ⁻ , CD64 ⁻ , CD3 ⁻ , NK1.1 ⁺ |
| NK-T cells | Viral and cancer immunity | CD45 ⁺ , F4/80 ⁻ , CD64 ⁻ , CD3 ⁺ , NK1.1 ⁺ |
| T cells | Immune regulation | CD45 ⁺ , F4/80 ⁻ , CD64 ⁻ , CD3 ⁺ , NK1.1 ⁻ |
| Macrophages | Phagocytosis and innate immunity | CD45 ⁺ , F4/80 ⁺ , CD64 ⁺ |
| cDC1 | Adaptive immunity; activation of CD8 ⁺ T cells | CD45 ⁺ , F4/80 ⁻ , CD64 ⁻ , CD3 ⁻ , NK1.1 ⁻ , CD19 ⁻ , MHCII ⁺ , CD11c ⁺ , CD172a ⁻ , XCR1 ⁺ |
| cDC2 | Humoral immunity; activation of CD4 ⁺ T cells | CD45 ⁺ , F4/80 ⁻ , CD64 ⁻ , CD3 ⁻ , NK1.1 ⁻ , CD19 ⁻ , MHCII ⁺ , CD11c ⁺ , CD172a ⁺ , XCR1 ⁻ |
| pDC | Viral immunity | CD45 ⁺ , F4/80 ⁻ , CD64 ⁻ , CD3 ⁻ , NK1.1 ⁻ , CD19 ⁻ , MHCII ⁺ , SiglecH ⁺ , CD11b ⁻ |

Table 1: Identification of different immune cell subsets in murine splenocytes using flow cytometry. Surface markers for characterization compiled from references 1, 2 and 3.

Results

Figure 1 depicts the gating strategy applied to identify the immune cell subsets of interest. By utilizing the expanded fluorescence capability of the MACSQuant Analyzer 16, it is possible to simultaneously detect the presence of T cell,

B cells, NK cells, NK-T cells, macrophages, conventional dendritic cells (cDCs) lineage 1, cDCs lineage 2 and plasmacytoid DCs (pDCs), as well as to screen for cell viability.

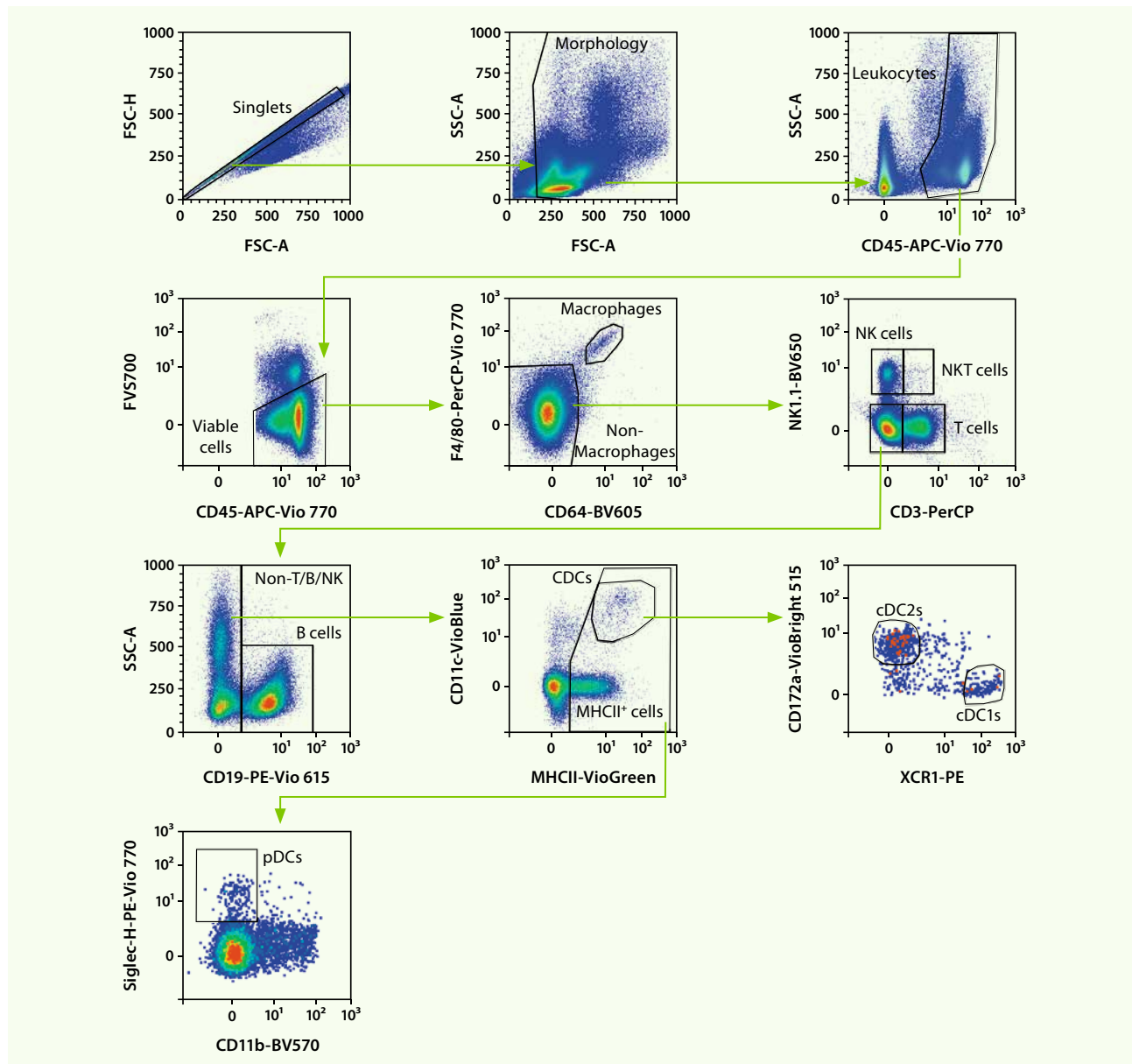


Figure 1: Immunophenotyping of mouse splenocytes. After doublet and dead cell exclusion, further gating was used to identify macrophages, T cells, NK cell, NKT cells, B cells, cDCs1, dDCs2 and pDCs.

Figure 2 shows the evaluation of the expression of the activation marker CD86 on all the above-identified subsets. The data clearly show, that the MACSQuant Analyzer 16 enables

high quality flow cytometry data acquisition with a 14-color panel, opening new possibilities for deeper phenotyping and immune monitoring studies.

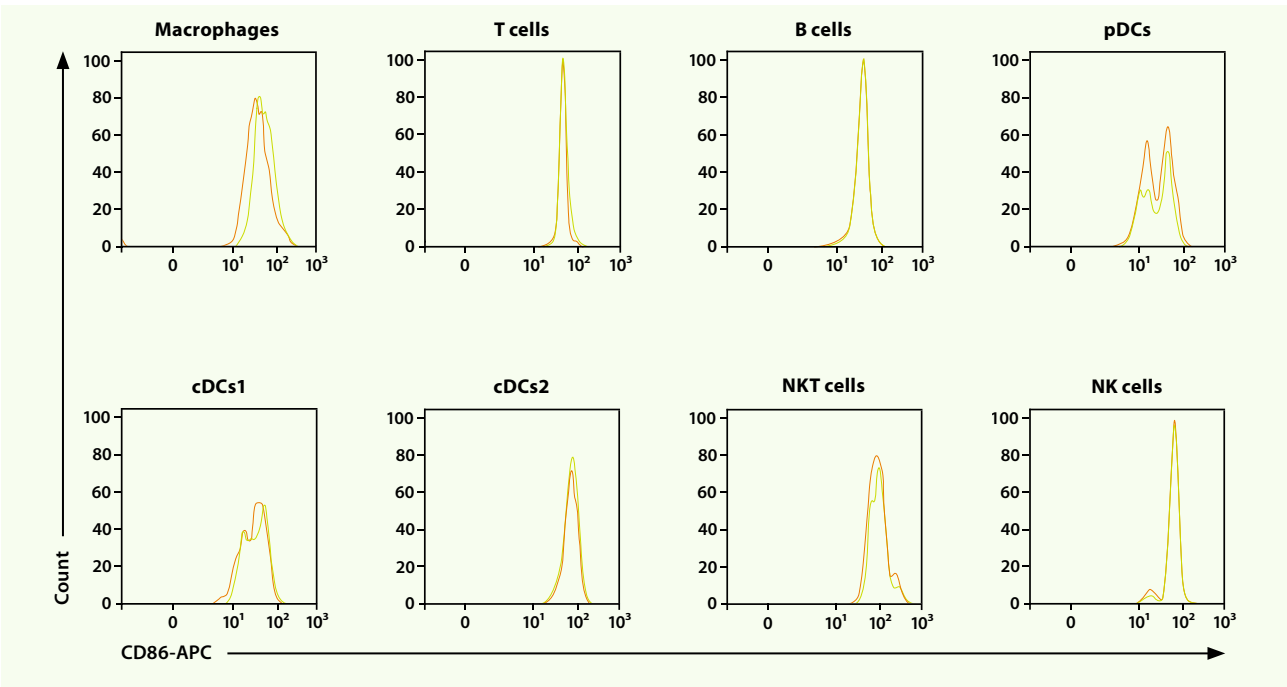


Figure 2: Expression of CD86 on different immune cell subsets. Staining for the cell activation marker CD86 (green) or appropriate control (Fluorescence minus one (FMO), red) on the different subsets identified in figure 1. Cave: staining performed on freshly isolated, unstimulated splenocytes.

Conclusion

This application note demonstrates the utility of flow cytometry for the detection and analysis of 8 immune cell subsets simultaneously including viability and activation status of the identified cells using a 14-color panel. It highlights:

- The possibility to design flow panels with up to 14 colors for the identification of different cellular subsets and for in-depth characterization of cells
- The advantage of high-content flow cytometric analyses when dealing with precious sample material
- The MACSQuant Analyzer 16 as a compact benchtop flow cytometer applicable for advanced immunophenotyping panels of murine cells

References

1. Martens *et al.* (2020) Nat. Immunol. 21, 381-387.
2. Gurka *et al.* (2015) Front. Immunol. 4:635.
3. Takagi *et al.* (2011) Immunity 35, 958-971

| Product | Clone | Order no. |
|--|-----------|-------------|
| Miltenyi Biotec products | | |
| CD11c Antibody, anti-mouse, VioBlue, REAfinity™ | REA754 | 130-110-706 |
| MHC Class II Antibody, anti-mouse, VioGreen, REAfinity | REA813 | 130-112-238 |
| CD172a (SIRPα) Antibody, anti-mouse, Vio Bright 515, REAfinity | REA1201 | 130-123-157 |
| XCR1 Antibody, anti-mouse/rat, PE, REAfinity | REA707 | 130-111-184 |
| CD19 Antibody, anti-mouse, PE-Vio 615, REAfinity™ | REA749 | 130-111-890 |
| F4/80 Antibody, anti-mouse, PerCP-Vio 770, REAfinity | REA126 | 130-118-327 |
| Siglec-H Antibody, anti-mouse, PE-Vio 770, REAfinity | REA819 | 130-112-140 |
| CD86 Antibody, anti-mouse, APC, REAfinity | REA1190 | 130-122-130 |
| CD45 Antibody, anti-mouse, APC-Vio 770, REAfinity | REA737 | 130-110-662 |
| Others | | |
| Brilliant Violet 570 anti-mouse/human CD11b Antibody | M1/70 | - |
| Brilliant Violet 605 anti-mouse CD64 Antibody | X54-5/7.1 | - |
| Brilliant Violet 650 anti-mouse NK1.1 Antibody | PK136 | - |
| PerCP anti-mouse CD3ε Antibody | 145-2C11 | - |
| BD Horizon™ Fixable Viability Stain (FVS) 700 | N/A | - |



Miltenyi Biotec

Miltenyi Biotec B.V. & Co. KG | Phone +49 2204 8306-0 | Fax +49 2204 85197 | macsde@miltenyi.com | www.miltenyibiotec.com

Miltenyi Biotec provides products and services worldwide. Visit www.miltenyibiotec.com/local to find your nearest Miltenyi Biotec contact.

Unless otherwise specifically indicated, Miltenyi Biotec products and services are for research use only and not for therapeutic or diagnostic use. MACSQuant, REAfinity, Vio, VioBlue, VioGreen, and the Miltenyi Biotec logo are registered trademarks or trademarks of Miltenyi Biotec and/or its affiliates in various countries worldwide. Copyright © 2022 Miltenyi Biotec and/or its affiliates. All rights reserved.