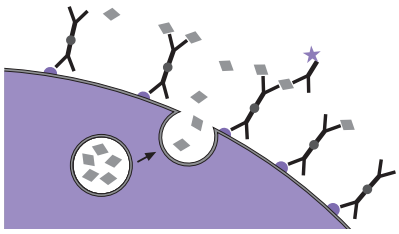
Miltenyi Biotec

# IL-5 Secretion Assay – Detection Kit (PE)

human

For 100 tests with $10^6$ cells

Order no. 130-091-623



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140-001-342.01



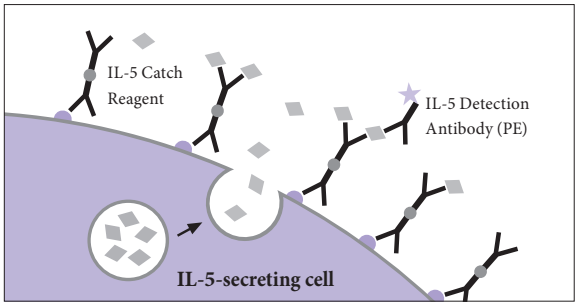


## Contents



## 1. Description

| | |
|---|---|
| **Components** | 1 mL **IL-5 Catch Reagent**: anti-IL-5 monoclonal antibody (rat IgG2a) conjugated to cell surface specific monoclonal antibody (mouse IgG2a).<br>1 mL **IL-5 Detection Antibody**: anti-IL-5 monoclonal antibody (rat IgG2a) conjugated to PE (R-phycoerythrin). |
| **Size** | For 100 tests with $10^6$ cells |
| **Product format** | IL-5 Catch Reagent and IL-5 Detection Antibody are supplied in a solution containing 0.1% gelatine and 0.05% sodium azide. |
| **Storage** | Store protected from light at 4–8 °C. Do not freeze. The expiration dates are indicated on the vial labels. |




## 1.1 Principle of the IL-5 Secretion Assay

Antigen-specific T cells are analyzed using the IL-5 Secretion Assay starting from whole blood, PBMCs or other leukocyte containing single-cell preparations. The cells are restimulated for a short period of time with specific peptide, protein or other antigen preparations.

Subsequently, an IL-5 specific **Catch Reagent** is attached to the cell surface of all leukocytes. The cells are then incubated for a short time at 37 °C to allow cytokine secretion. The secreted IL-5 binds to the IL-5 Catch Reagent on the positive, secreting cells. These cells are subsequently labeled with a second IL-5-specific antibody, the **IL-5 Detection Antibody** conjugated to R-phycoerythrin (PE) for sensitive detection by flow cytometry. Since viable cells are analyzed, non-specific background can be minimized by dead cell exclusion. This provides highest sensitivity of analysis.



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## 1.2 Background and product applications

The IL-5 Secretion Assay - Detection Kit is designed for the detection and analysis of viable IL-5-secreting leukocytes. It is specially developed for the **detection of antigen-specific T cells**. After restimulation with specific antigen *in vitro* secretion of IL-5 is induced.

IL-5 (interleukin 5) is a cytokine predominantly secreted by $CD4^+$ T cells. It is involved in a range of allergic reactions and mediates immune reactions against parasites. IL-5 also acts on other cell types, like B cells.

Quantitative analysis of antigen-specific T cell populations can provide important information on the natural course of immune responses.

**Examples of applications**

- Detection of viable IL-5-secreting leukocytes.
- Detection of IL-5-secreting, antigen-specific T cells for enumeration and phenotypic characterization.
- Monitoring and analysis of antigen-specific T cell immunity, e.g. in infection, autoimmunity, cancer, allergy or alloreactivity.
- Analysis of IL-5-secreting cells for determination of functional antigens in disease and for T cell receptor (TCR) epitope mapping.
- Analysis of TCR repertoire of antigen-specific T cells.

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## 1.3 Reagent and instrument requirements

- **Buffer** (degassed): Prepare a solution containing PBS (phosphate buffered saline) pH 7.2, 0.5% BSA and 2 mM EDTA by diluting MACS BSA Stock Solution (# 130-091-376) 1:20 with autoMACS™ Rinsing Solution (# 130-091-222). Keep buffer cold (4–8 °C).
- **Culture medium**, e.g. RPMI 1640 (# 130-091-440) containing 5% human serum, like autologous or AB serum (do **not use BSA** or **FCS** because of non-specific stimulation!).
- **Propidium iodide (PI)** or **7-AAD** for flow cytometric exclusion of dead cells. For cell fixation and flow cytometric exclusion of dead cells, the **Fixation and Dead Cell Discrimination Kit** (#130-091-163) is recommended.
- Refrigerated centrifuge (4–8 °C).
- Rotation device for tubes: MACSmix™ tube rotator (# 130-090-753).
- (Optinal) CytoStim (# 130-092-172), reagent for stimulation of T cells.
- (Optional) Staining reagents such as CD4-FITC (# 130-080-501) or CD8-FITC (# 130-080-601) and CD14-PerCP.



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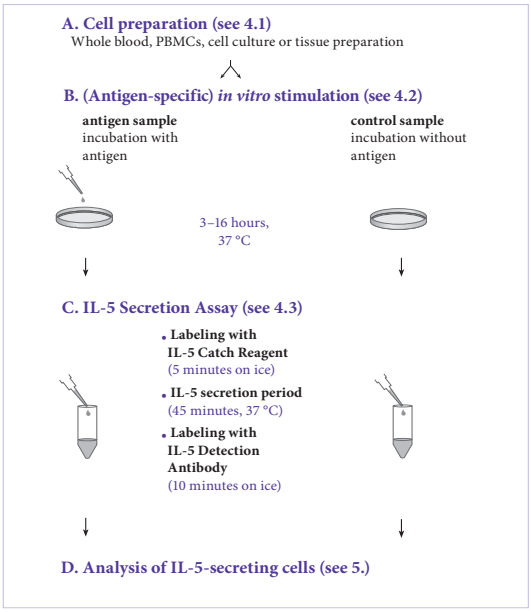
# 2. Protocol overview

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# 3. Experimental set-up

## 3.1 Controls

**Negative control**
For accurate detection of IL-5-secreting antigen-specific cells, a negative control sample should always be included. This will provide information about IL-5 secretion unrelated to the specific antigen-stimulation, but e.g. due to ongoing in vivo immune responses. The control sample should be treated exactly the same as the antigen-stimulated sample except for the addition of antigen, or by using a control antigen.

**Positive control**
When setting up a new experiment, it is recommended to include a positive control. As a positive control, a sample stimulated with CytoStim (# 130-092-172) 20 µL/mL for 1–3 hours, or the superantigen Staphylococcal Enterotoxin B (Sigma) 1 µg/mL for 3–16 hours, may be included in the experiment.

## 3.2 Kinetics of restimulation and proposed time schedule

**Peptides**
Upon stimulation with peptide, the cells can be analyzed for IL-5 secretion 3–6 hours later.
It is possible to prepare the cells first and take them into culture overnight, but without adding the antigen (see 4.2 step 2.). Peptide is then added the next morning for 3 hours of stimulation, directly followed by the IL-5 Secretion Assay.





**Proteins**
Upon stimulation with protein, the cells can be analyzed for IL-5 secretion 6-16 hours later.
It is possible to start the stimulation of the cells late in the afternoon, and to perform the IL-5 Secretion Assay the following morning.

**Co-stimulation**
The addition of co-stimulatory agents like CD28 or CD49d antibody may enhance the response to the antigen. If co-stimulatory agents are added to the antigen sample, they also have to be included in the control sample.

## 3.3 Counterstaining of cytokine-secreting cells

The IL-5-secreting cells are stained with PE-conjugated IL-5 Detection Antibodies. To identify cells of interest, counterstaining for T cells with e.g. CD4-FITC (# 130-080-501) or CD8-FITC (# 130-080-601) is important.

▲ Upon activation of T cells, TCR and some associated molecules, like CD3, might be down-regulated.

▲ The samples should be stained with propidium iodide (PI) or 7-AAD prior to acquisition, to exclude dead cells from analysis. This will reduce non-specific background staining and increase sensitivity.

▲ For optimal sensitivity, we recommend labeling of undesired non-T cells such as monocytes with antibodies conjugated to PerCP, e.g. CD14-PerCP. These cells can then be excluded together with PI stained dead cells by gating.





## 3.4 Two color cytokine analysis (see also 5.1)

IL-5-secreting cells can be analyzed simultaneously for IFN-γ, IL-2 IL-10, or TNF-α production by two color cytokine analysis combining the IL-5 Secretion Assay with the IFN-γ Secretion Assay - Detection Kit (APC) (# 130-090-762), the IL-2 Secretion Assay - Detection Kit (APC) (# 130-090-763), the IL-10 Secretion Assay- Detection Kit (APC) (# 130-090-761), or the TNF-α Secretion Assay- Detection Kit (APC) (# 130-091-624). Detailed protocols are included in the data sheets of the Cytokine Secretion Assay - Detection Kits (APC) and are available from our website www.miltenyibiotec.com/protocols.

## 3.5 Combination with peptide-MHC tetramer staining

IL-5-secreting cells can be analyzed simultaneously for peptide-MHC tetramers combining the IL-5 Secretion Assay (PE) with APC-conjugated peptide-MHC tetramers. For combination with PE-conjugated peptide-MHC tetramers the IL-5 Secretion Assay - Detection Kit (APC) (# 130-091-624) is available. Detailed recommendations for the experimental set-up and the procedure are included in the data sheets of the Cytokine Secretion Assay - Detection Kits (APC) and are available from our website www.miltenyibiotec.com/protocols.

## 3.6 Detection of very low frequencies

(Optional, reagents not included) If the sample contains fewer





than 0.01–0.1% of IL-5-secreting cells, it is possible to enrich these cells magnetically using the IL-5 Secretion Assay – Enrichment and Detection Kit (# 130-091-622). Thereby it is possible to detect antigen-specific T cells down to frequencies as low as 0.0001% (1 in $10^6$).

# 4. Protocol for the IL-5 Secretion Assay

## 4.1 Cell preparation

For the detection of cytokine-secreting cells, best results are achieved by starting the assay with fresh PBMCs, or other leukocyte containing single-cell preparations from tissues or cell lines. Alternatively, frozen cell preparations can be used.

▲ **Note:** PBMCs may be stored over night. The cells should be resuspended and incubated in culture medium as described in 4.2 step 2., but without addition of antigen. The antigen is then added to the culture on the next day.

▲ **Note:** Remove platelets after density gradient separation. Resuspend cell pellet, fill tube with buffer and mix. Centrifuge at 200×g for 10–15 minutes at 20 °C. Carefully remove supernatant.

**Special protocols for whole blood:** You can start the IL-5 Secretion Assay directly from whole blood. For details on the procedure, see 7. Appendix B: **Detection of cytokine-secreting cells from human whole blood.** This special protocol is also available from our website www.miltenyibiotec.com/protocols.

## 4.2 (Antigen specific) *in vitro* stimulation

▲ Always include a **negative control** in the experiment. A **positive control** may also be included (see 3.1).

▲ Do **not use** media containing any **non-human** proteins, like BSA or FCS because of non-specific stimulation.

**Protocol for *in vitro* stimulation**

1. Wash cells by adding medium, centrifuge at 300×g for 10 minutes.
2. Resuspend cells in culture medium, containing 5% human serum, adjust to $10^7$ cells/mL and $5\times10^6$ cells/cm² (see 7. Appendix A: Flask and dish sizes for stimulation).
3. Add antigen or control reagent:

   peptide: 3–6 hours at 37 °C, 5–7% $CO_2$, e.g. 1–10 µg/mL
   protein: 6–16 hours at 37 °C, 5–7% $CO_2$, e.g. 10 µg/mL
   CytoStim: 1–3 hours at 37 °C, 5–7% $CO_2$, 20 µL/mL
   SEB: 3–16 hours at 37 °C, 5–7% $CO_2$, e.g. 1 µg/mL

   For comparison of different experiments, the stimulation time should always be the same (see 3.2).
4. Collect cells carefully by using a cell scraper, or by pipetting up and down when working with smaller volumes. Rinse the dish with cold buffer. Check microscopically for any remaining cells, if necessary, rinse the dish again.





## 4.3 Cytokine Secretion Assay

### General considerations

▲ The assay is optimized for cell samples containing < 5% of total IL-5-secreting cells. If ≥ 5% of IL-5-secreting cells are expected, it is necessary to dilute the cells further during the cytokine secretion period, and therefore a larger test tube will be needed (see table below). The dilution prevents non-specific staining of cells not secreting IL-5 during this period.

▲ For each test with $10^6$ total cells, prepare:

50 mL of **cold buffer** (4–8 °C)
100 µL of **cold medium** (4–8 °C)
1 mL (or 10 mL; see table below) of **warm medium** (37 °C).

▲ Work fast, keep the cells cold, use pre-cooled solutions which will prevent capping of antibodies on the cell surface and a non-specific cell labeling (exception: warm medium during secretion period).

▲ Volumes shown below are for $10^6$ total cells. When working with fewer than $10^6$ cells, use the same volumes as indicated. When working with higher cell numbers, scale up all reagent volumes and total volumes, accordingly (e.g. for $2\times10^6$ total cells, use twice the volume of all indicated reagent volumes and total volumes).

▲ Do not remove supernatant by decanting. This will lead to cell loss and incorrect incubation volumes. Pipette off or aspirate supernatant.





**Labeling cells with IL-5 Catch Reagent**

1. Use $10^6$ total cells in a 2 mL closable tube per sample.

   ▲ **Note:** For larger cell numbers, scale up all volumes accordingly. For fewer than $10^6$ cells, use same volumes.
2. Wash cells by adding 1–2 mL of **cold buffer**, centrifuge at 300×g for 10 minutes at **4–8 °C**, pipette off supernatant completely.

   ▲ **Note**: Do not remove supernatant by decanting. This will lead to cell loss and incorrect incubation volumes.
3. Resuspend cell pellet in 90 µL of **cold medium** per $10^6$ total cells.
4. Add 10 µL of **IL-5 Catch Reagent** per $10^6$ total cells, mix well and incubate for 5 minutes **on ice**.

**IL-2 secretion period**

1. Add **warm** (37 °C) medium to dilute the cells according to the following table:

| Expected number of IL-5-secreting cells | Dilution | Amount of medium to add per $10^6$ total cells |
|---|---|---|
| < 5 % | $10^6$ cells/mL | 1 mL |
| ≥ 5 % | ≤ $10^5$ cells/mL | 10 mL |

   ▲ **Note:** For frequencies of cytokine-secreting cells >> 20% the cells need to be further diluted, e.g. by a factor of 5.
2. Incubate cells in closed tube for 45 minutes at 37 °C under slow continuous rotation using the MACSmix™ tube rotator (# 130-090-753), or turn tube every 5 minutes to resuspend settled cells.





▲ **Note:** During this step it is crucial to prevent contact of cells to avoid cross contamination with cytokines.

**Labeling cells with IL-5 Detection Antibody**

1. Put the tube **on ice**.
2. Wash the cells by filling up the tube with **cold buffer**, centrifuge at 300×g for 10 minutes at **4–8 °C**. Pipette off supernatant completely.

   ▲ **Note**: If the volume of the cell suspension was higher than the volume of added buffer, repeat wash step.
3. Resuspend cell pellet in 90 µL of **cold buffer** per $10^6$ total cells.
4. Add 10 µL of **IL-5 Detection Antibody (PE)** per $10^6$ total cells.
5. (Optional) Add additional staining reagents, e.g. 10 µL of CD4-FITC (# 130-080-501) or 10 µL of CD8-FITC (# 130-080-601) and CD14-PerCP™.
6. Mix well and incubate for 10 minutes **on ice**.
7. Wash cells by adding 2 mL of **cold buffer**, centrifuge at 300×g for 10 minutes at **4–8 °C**, pipette off supernatant.
8. Proceed to analysis (see section 5.).

## 5. Detection and analysis of IL-5-secreting T cells

▲ Add propidium iodide (PI) or 7-AAD to a final concentration of 0.5 µg/mL **just prior** to acquisition for exclusion of dead cells from flow cytometric analysis. Incubating with PI for longer periods will affect the viability of the cells.
**Do not fix the cells** when using PI or 7-AAD.

▲ For optimized sensitivity, an appropriate number of viable cells has to be acquired from the antigen stimulated sample as well as from the control sample.

- Acquire $2\times10^5$ viable cells from each sample.

To illustrate the analysis, we describe the detection of IL-5-secreting T cells using the IL-5 Secretion Assay. The detailed description, including how to set gates, should serve as a model for the analysis of your own sample.

1. $10^7$ human PBMCs were restimulated for 1 hour with and without CytoStim (# 130-092-172).
2. The IL-5 Secretion Assay was performed on the stimulated and the unstimulated sample.
3. **Counterstaining of T cells** was performed using CD4-FITC.
4. **Dead cells** were stained with propidium iodide (PI), which was added just prior to flow cytometric analysis to a final concentration of 0.5 µg/mL.



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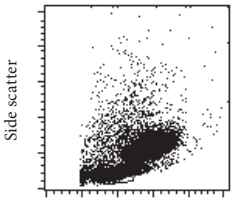
5. Detection and analysis of IL-5-secreting T cells

5. 200,000 viable cells were acquired by flow cytometry, from the stimulated as well as from the unstimulated sample.
6. A **lymphocyte gate** based on forward and side scatter (FSC/SSC) properties was activated prior to further gating to exclude monocytes and debris (see A.).
7. Dead cells and monocytes were excluded according to PI staining in a fluorescence 2 (PE) versus fluorescence 3 plot (see B.).

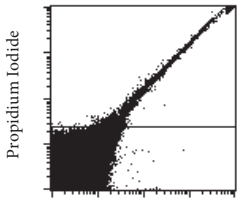
- The **dead cell exclusion** is crucial for the analysis of rare antigen-specific T cells, as dead cells may bind non-specifically to antibodies or MicroBeads. This could lead to false positive events.

8. Analysis of secreted IL-5 (PE) versus CD4-FITC staining of viable lymphocytes is displayed (see C.).

**A. Lymphocyte gate using FSC versus SSC**




**B. Dead cell exclusion**



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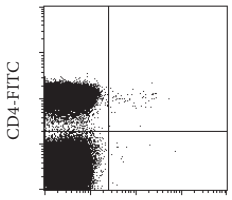
**C. IL-5-secreting CD4⁺ T cells after stimulation with CytoStim**

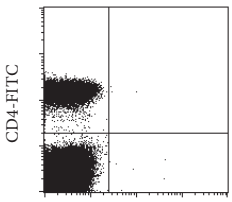
**Sample stimulated with CytoStim**




0.083% of the total CD4⁺ T cell population secrete IL-5 (see formula below).

$$\%\ \text{IL-5}^+\ \text{cells among CD4}^+ = \frac{\#\ \text{of IL-5}^+\text{CD4}^+\ \text{cells in the analyzed sample}}{\#\ \text{of total CD4}^+\ \text{cells in the analyzed sample}} \times 100$$

**Unstimulated control sample**




≤ 0.008% of the total CD4⁺ T cell population secrete IL-5.



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6. References

## 6. References

1. Manz, R; Assenmacher, M; Pflüger, E; Miltenyi, S; Radbruch, A (1995) Analysis and Sorting of Live cells According to Secreted Molecules Relocated to a Cell-Surface Affinity Matrix. Proc.Natl.Acad.Sci. USA 92: 1921-1925. [139]
2. Assenmacher, M; Löhning, M; Scheffold, A; Manz, RA; Schmitz, J; Radbruch, A. (1998) Sequential production of IL-2, IFN-γ and IL-10 by individual staphylococcal enterotoxin B-activated T helper lymphocytes. Eur. J. Immunol. 28: 1534-1543. [483]
3. Brosterhus, H; Brings, S; Leyendeckers, H; Manz, RA; Miltenyi, S; Radbruch, A; Assenmacher, M; Schmitz, J (1999) Enrichment and detection of live antigen-specific CD4⁺ and CD8⁺ T cells based on cytokine secretion. Eur. J. Immunol. 29: 4053-4059. [573]
4. Oelke, M; Moehrle, U; Chen, JL; Behringer, D; Cerundolo, V; Lindemann, A; Mackensen, A (2000) Generation and purification of CD8⁺ Melan-A-Specific Cytotoxic T Lymphocytes for Adoptive Transfer in Tumor Immunotherapy. Clin. Cancer Res. 6: 1997-2005. [663]
5. Oelke, M; Kurokawa,T; Hentrich, I.; Behringer, D; Cerundolo, V; Lindemann, A; Mackensen, A (2000) Functional Characterization of CD8⁺ Antigen-Specific Cytotoxic T Lymphocytes after Enrichment Based on Cytokine Secretion: Comparison with the MHC-Tetramer Technology. Scand. J. Immunol. 52. 544-549. [970]
6. Bickham, K; Münz, C; Tsang, ML; Larsson, M; Fonteneau, J-F; Bhardwaj, N; Steinmann, R (2001) EBNA1-specific CD4⁺T cells in healthy carriers of Epstein-Barr virus are primarily Th1 in function. J. Clin. Invest. 107: 121-130. [1035]
7. Pittet, MJ; Zippelius, A; Speiser, DE; Assenmacher, M; Guillaume, P; Valmori, D; Lienard, D; Lejeune, F; Cerottini, JC; Romero, P (2001) Ex vivo IFN-γ secretion by circulating CD8 T lymphocytes: Implications of a novel approach for T cell monitoring in infectious malignant diseases. J. Immunol. 166: 7634-7640. [1037]
8. Becker, C; Pohla, H; Frankenberger, F; Schüler, T; Assenmacher, M; Schendel, DJ; Blankenstein, T (2001) Adoptive tumor therapy with T lymphocytes enriched through an IFN-γ capture assay. Nature Medicine 7, 10: 1159-1162. [1207]

For further references visit our website **www.miltenyibiotec.com**.



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# 7. Appendix

## A: Flask and dish sizes for stimulation

For *in vitro* stimulation (see 4.2 step 2.) the cells should be resuspended in culture medium, containing 5% of human serum, at $10^7$ cells/mL and $5\times10^6$ cells/cm$^2$. Both the dilution and the cell density are important to assure optimum stimulation.

The following table lists culture plate, dish and flask sizes suitable for different cell numbers. It also indicates the appropriate amount of medium to add.

| total cell number | medium volume to add | culture plate | well diameter |
|---|---|---|---|
| $0.15\times10^7$ | 0.15 mL | 96 well | 0.64 cm |
| $0.5\times10^7$ | 0.5 mL | 48 well | 1.13 cm |
| $1\times10^7$ | 1 mL | 24 well | 1.6 cm |
| $2\times10^7$ | 2 mL | 12 well | 2.26 cm |
| $5\times10^7$ | 5 mL | 6 well | 3.5 cm |

| total cell number | medium volume to add | culture dish | dish diameter |
|---|---|---|---|
| $4.5\times10^7$ | 4.5 mL | small | 3.5 cm |
| $10\times10^7$ | 10 mL | medium | 6 cm |
| $25\times10^7$ | 25 mL | large | 10 cm |
| $50\times10^7$ | 50 mL | extra large | 15 cm |

| total cell number | medium volume to add | culture flask | growth area |
|---|---|---|---|
| $12\times10^7$ | 12 mL | 50 mL | 25 cm$^2$ |
| $40\times10^7$ | 40 mL | 250 mL | 75 cm$^2$ |
| $80\times10^7$ | 80 mL | 720 mL | 162 cm$^2$ |
| $120\times10^7$ | 120 mL | 900 mL | 225 cm$^2$ |





## B: Detection of cytokine-secreting cells from whole blood

**B1. Reagent and instrument requirements**

**B2. Protocol**

**B 2.1 (Antigen-specific) *in vitro* stimulation**

**B 2.2 Cytokine Secretion Assay**

**B 2.3 Detection and analysis of cytokine-secreting cells**

**The following special protocol can be used in combination with one of the Cytokine Secretion Assay - Detection Kits for human cells.**

## B 1. Reagent and instrument requirements

- **Cytokine Secretion Assay, for example:**

| | |
|---|---|
| IFN-γ Secretion Assay - Detection Kit (PE) | (# 130-054-202) |
| IFN-γ Secretion Assay - Detection Kit (FITC) | (# 130-090-433) |
| IFN-γ Secretion Assay - Detection Kit (APC) | (# 130-090-762) |
| IL-2 Secretion Assay - Detection Kit (PE) | (# 130-090-487) |
| IL-2 Secretion Assay - Detection Kit (APC) | (# 130-090-763) |
| IL-4 Secretion Assay - Detection Kit (PE) | (# 130-054-102) |
| IL-5 Secretion Assay - Detection Kit (PE) | (# 130-091-623) |
| IL-5 Secretion Assay - Detection Kit (APC) | (# 130-091-624) |
| IL-10 Secretion Assay - Detection Kit (PE) | (# 130-090-434) |
| IL-10 Secretion Assay - Detection Kit (APC) | (# 130-090-761) |
| TNF-α Secretion Assay - Detection Kit (PE) | (# 130-091-268) |
| TNF-α Secretion Assay - Detection Kit (APC) | (# 130-091-267) |

- **Anticoagulant:** sodium heparin





- **Buffer** (degassed): Prepare a solution containing PBS (phosphate buffered saline) pH 7.2, 0.5% BSA and 2 mM EDTA by diluting MACS BSA Stock Solution (# 130-091-376) 1:20 with autoMACS™ Rinsing Solution (# 130-091-222). Keep buffer cold (4–8 °C).
- **Culture medium**, e.g. RPMI 1640 containing 10% of human serum, like autologous serum or AB serum.

  ▲ **Note:** Do **not use BSA** or **FCS** because of non-specific stimulation.
- (Optinal) CytoStim™ (# 130-092-172), reagent for stimulation of T cells.
- **Erythrocyte lysing solution (1×):**

- prepare freshly from 10× stock solution.
- **10× stock solution**: 41.4 g $NH_4Cl$ (1.55 M), 5 g $KHCO_3$ (100 mM), 1 mL 0.5 M EDTA (1 mM), adjust pH to 7.3, fill up to 500 mL with dd $H_2O$.

  ▲ **Note:** Do **not use** FACS Lysing solution™.

- (Optional) **Staining reagents**: CD4-FITC (# 130-080-501) or CD8-FITC (# 130-080-601) and CD14-PerCP™.

  ▲ **Note:** Upon activation of T cells, TCR and some associated molecules, like CD3, might be down-regulated.

  ▲ **Note**: For optimal sensitivity, we recommend labeling of undesired non-T cells such as monocytes with antibodies conjugated to PerCP™, e.g. CD14-PerCP™. These cells can then be excluded together with PI stained dead cells by gating.
- **Propidium iodide (PI)** or **7-AAD** for flow cytometric exclusion of dead cells. For cell fixation and flow cytometric exclusion of dead cells, the **Fixation and Dead Cell Discrimination Kit** (#130-091-163) is recommended.
- (Optional) Rotation device for tubes: MACSmix™ tube rotator (# 130-090-753).





## B 2. Protocol

▲ The peripheral blood should not be older than 20 hours and should be supplemented with anticoagulant **sodium heparin**. **Do not use EDTA or ACD**. Lymphocyte activation and secretion of cytokines requires calcium, and is consequently inhibited by chelating anticoagulants.

▲ **Note**: Whole blood may be stored over night at room temperature.

▲ Always include a **negative control** sample in the experiment. A **positive control** with e.g. CytoStim (# 130-092-172), or Staphylococcal Enterotoxin B (SEB) may be included in the experiment (see also detailed protocol provided with the Cytokine Secretion Assay Kits).

▲ Do **not use** media containing any **non-human** proteins, like BSA or FCS because of non-specific stimulation.

**Protocol for *in vitro* stimulation**

1. Start with **250 µL of fresh, sodium heparinized, human blood** (containing about $5\times10^5$ lymphocytes) in a 15 mL conical polypropylene tube.
2. Add the antigen or, as a positive control, 1 µg/mL SEB for 3–16 hours at 37 °C, 5-7% $CO_2$ (for details on the kinetics of cytokine

secretion and on concentrations of antigen to add, refer to Cytokine Secretion Assay data sheet, 3.1-3.2).

3. A negative control sample, treated exactly the same as the antigen-stimulated sample, but without addition of antigen, should always be included in the experiment.
4. (Optional) Co-stimulatory agents like CD28 and CD49d antibodies may be added.

### B 2.2 Cytokine Secretion Assay

▲ This protocol is optimized for cell samples containing < 20% of total cytokine-secreting cells. If ≥ 20% of cytokine-secreting cells are expected, it is necessary to dilute the cells further during the cytokine secretion period, and therefore a larger test tube will be needed. The dilution prevents non-specific staining of cells not secreting cytokines during this period.

▲ For each sample with 250 µL whole blood prepare:

50 mL of **cold buffer** (4–8 °C)
100 µL of **cold medium** (4–8 °C)
5 mL of **warm medium** (37 °C)
5 mL of **erythrocyte lysing solution** (room temperature).

▲ Work fast, keep the cells cold, use pre-cooled solutions which will prevent capping of antibodies on the cell surface and a non-specific cell labeling (exception: warm medium during secretion period and room temperature during lysing step).



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▲ Do not remove supernatant by decanting. This will lead to cell loss and incorrect incubation volumes. Pipette off or aspirate supernatant.

▲ Dead cells may bind non-specifically to MACS MicroBeads or antibodies. Therefore, when working with cell preparations containing large amounts of dead cells, they should be removed before starting the Cytokine Secretion Assay, e.g. by density gradient centrifugation or by using the Dead Cell Removal Kit (# 130-090-101).

#### Labeling cells with Cytokine Catch Reagent

▲ Higher temperatures and longer incubation times for staining should be avoided. This will lead to non-specific cell labeling.

1. Wash cells by adding 10 mL of **cold buffer**, centrifuge at 300×g for 10 minutes at 4–8 °C, pipette off supernatant carefully.
   ▲ **Note**: Be careful, leukocytes will appear on top of the loose red cell pellet.
2. Resuspend pellet in 80 µL of **cold medium**.
3. Add 20 µL of **Cytokine Catch Reagent**, mix well and incubate for 5 minutes **on ice**.

#### Cytokine secretion period

1. Add 5 mL of **warm medium** (37 °C) to dilute the cells.
   ▲ **Note:** For frequencies of cytokine-secreting cells ≥ 20% the cells need to be further diluted, e.g. by a factor of 5.



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2. Incubate cells in closed tube for 45 minutes at **37 °C** under slow continuous rotation by using the MACSmix™ tube rotator (# 130-090-753), or turn tube every 5 minutes to resuspend settled cells.
   ▲ **Note:** During this step it is crucial to prevent contact of cells to avoid cross contamination with cytokines.

#### Labeling cells with Cytokine Detection Antibody

1. Put the tube **on ice**.
2. Wash cells by adding 10 mL of **cold buffer**, centrifuge at 300×g for 10 minutes at **4–8 °C**, pipette off supernatant carefully.
3. Resuspend cell pellet in 80 µL of **cold buffer**.
4. Add 20 µL of **Cytokine Detection Antibody**.
5. (Optional) Add additional staining reagents, e.g. 10 µL of CD4-FITC (# 130-080-501) or CD8-FITC (# 130-080-601) and CD14-PerCP™.
6. Mix well and incubate for 10 minutes **on ice**.

#### Lysis of erythrocytes

1. Add 5 mL of **erythrocyte lysing solution**.
2. Mix gently and incubate for 10 minutes at room temperature. Rotate tube continuously using the MACSmix tube rotator, or turn tube several times during incubation.
3. Centrifuge cells at 300×g for 10 minutes at **room temperature**, pipette off supernatant completely.
4. Wash cells by adding 10 mL of **cold buffer**, centrifuge at 300×g for 10 minutes at 4–8 °C, pipette off supernatant.
5. Resuspend the cells in 500 µL of **cold buffer**, and proceed to flow cytometric analysis (see detailed protocol).

### B 2.3 Detection and analysis of cytokine-secreting cells

▲ Add propidium iodide (PI) or 7-AAD to a final concentration of 0.5 µg/mL **just prior** to acquisition to exclude dead cells from flow cytometric analysis. Incubation with PI for longer periods will affect the viability of the cells.
**Do not fix the cells** when using PI or 7-AAD.

▲ For optimized sensitivity, an appropriate number of viable cells has to be acquired from the antigen **stimulated sample** as well as from the **control sample**.

- Acquire $2\times10^5$ viable cells from each sample.

▲ For details on analysis please refer to the detailed protocol provided with the Cytokine Secretion Assay Kits.

Refer to **www.miltenyibiotec.com** for all data sheets and protocols. Miltenyi Biotec provides technical support worldwide. Visit www.miltenyibiotec.com for local Miltenyi Biotec Technical Support contact information.

### Warnings

Reagents contain sodium azide. Under acidic conditions sodium azide yields hydrazoic acid, which is extremely toxic. Azide compounds should be diluted with running water before discarding. These precautions are recommended to avoid deposits in plumbing where explosive conditions may develop.

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