



Miltenyi Biotec

Reference list

T Cell TransAct™ – Next-generation T cell activation and expansion reagent

What is T Cell TransAct?

T Cell TransAct is a ready-to-use reagent for physiological activation and expansion of human T cells.

- The colloidal polymeric nanomatrix conjugated to humanized recombinant CD3 and CD28 agonists mimics *in vivo* T cell activation.
- Ensures robust activation of resting T cells from haematological cell populations (e.g. PBMCs or enriched T cell populations) without the involvement of CD4 or CD8.
- Easy protocol with no need for cell counting, plate coating, or bead removal steps. The reagent is simply washed off.
- Available as research use only (RUO) product and in MACS® GMP Quality.
- MACS GMP T Cell TransAct is compatible with the CliniMACS Prodigy® Platform for automated cell manufacturing in a closed system.

Selected references

T Cell TransAct Reagent is successfully applied in various application fields worldwide. In the following, we have compiled a selection of recent high-impact publications using the RUO product.

Cell and gene therapy research

Autologous T cell immunotherapy model produced in a stirred tank bioreactor system using T Cell TransAct.

Baradez, M. O. *et al.* (2018) Application of Raman spectroscopy and univariate modelling as a process analytical technology for cell therapy bioprocessing. *Front. Med.* 5: 47. doi: 10.3389/fmed.2018.00047. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5844923/>

T cell activation to increase transduction efficiency during the production of lentiviral vectors.

Bauler, M. *et al.* (2020) Production of lentiviral vectors using suspension cells grown in serum-free media. *Mol. Ther. Methods Clin. Dev.* 17: 58–68. doi: 10.1016/j.omtm.2019.11.011. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6931067/>

Transduction of T Cell TransAct-activated CD8⁺ T cells with TCR_{MART-1} lentivirus.

Ding, R. *et al.* (2021) Single-cell transcriptome analysis of the heterogeneous effects of differential expression of tumor PD-L1 on responding TCR-T cells. *Theranostics* 11: 4957–4974. doi: 10.7150/thno.5507. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7978322/>

Establishment of a novel humanized artificial receptor platform termed RevCARs for CAR T cell optimization. Pan T cell activation using TransAct.

Feldmann, A. *et al.* (2020) Versatile chimeric antigen receptor platform for controllable and combinatorial T cell therapy. *Oncoimmunology* 9: 1785608. doi: 10.1080/2162402X.2020.1785608. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7458653/>

Generation and functional validation of NKG2DIL7-CAR T cells.

He, C. *et al.* (2020) Co-expression of IL-7 improves NKG2D-based CAR T cell therapy on prostate cancer by enhancing the expansion and inhibiting the apoptosis and exhaustion. *Cancers* 12: 1969. doi: 10.3390/cancers12071969. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7409228/>

T cell isolation and generation of adapter CAR T cells for ovarian carcinoma using T Cell TransAct, TexMACS™ Medium, and MACS Cytokines.

Kinkhabwala A. *et al.* (2022) MACSima imaging cyclic staining (MICS) technology reveals combinatorial target pairs for CAR T cell treatment of solid tumors. *Sci. Rep.* 12: 1911. doi: 10.1038/s41598-022-05841-4. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8813936/>

Optimized delivery of mRNA into primary human T cells by an innovative microfluidic continuous-flow electrotransfection device designed for precise, consistent, and high-throughput genetic modification of target cells in cellular therapy manufacturing applications.

Lissandrello, C. A. *et al.* (2020) High-throughput continuous-flow microfluidic electroporation of mRNA into primary human T cells for applications in cellular therapy manufacturing. *Sci. Rep.* 10: 18045. doi: 10.1038/s41598-020-73755-0. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7582186/>

T Cell TransAct–induced pan T cell activation for the generation of CAR T cells.

Niño, J. L. G. *et al.* (2020) Cytotoxic T cells swarm by homotypic chemokine signalling. *eLife* 9: e56554. doi: 10.7554/eLife.56554. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7669268/>

Lentiviral transduction of PBMCs previously activated with T Cell TransAct.

Oh, J. *et al.* (2019) Single variable domains from the T cell receptor β chain function as mono- and bifunctional CARs and TCRs. *Sci. Rep.* 9: 17291. doi: 10.1038/s41598-019-53756-4. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6872726/>

Generation and validation of CD123-CAR^{CD20} T cells.

Riberdy, J. M. *et al.* (2020) The art and science of selecting a CD123-specific chimeric antigen receptor for clinical testing. *Mol. Ther. Methods Clin. Dev.* 18: 571–581. doi: 10.1016/j.omtm.2020.06.024. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7393323/>

CRISPR-Cas9-based genome editing of human primary pan T cells isolation with the Pan T Cell Isolation Kit and activated using T Cell TransAct, TexMACS Medium, and MACS Cytokines.

Robert, P. *et al.* (2021) Functional mapping of adhesiveness on live cells reveals how guidance phenotypes can emerge from complex spatiotemporal integrin regulation. *Front. Bioeng. Biotechnol.* 9: 625366. doi: 10.3389/fbioe.2021.625366. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8058417/>

Small-scale CAR T cell generation using T Cell TransAct, TexMACS Medium, and IL-2.

Schäfer, D. *et al.* (2020) A novel siglec-4 derived spacer improves the functionality of CAR T cells against membrane-proximal epitopes. *Front. Immunol.* 11: 1704. doi: 10.3389/fimmu.2020.01704. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7426717/>

CAR T cell generation and functional validation of newly identified CAR T cell targets.

Schäfer, D. *et al.* (2021) Identification of CD318, TSPAN8, and CD66c as target candidates for CAR T cell–based immunotherapy of pancreatic adenocarcinoma. *Nat. Commun.* 12: 1453. doi: 10.1038/s41467-021-21774-4. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7935963/>

Novel system to simplify application of TRUCK (“T cells redirected for universal cytokine-mediated killing”)–modified T cells in solid tumor therapy.

Zimmermann, K. *et al.* (2020) Design and characterization of an “all-in-one” lentiviral vector system combining constitutive anti-G_{D2} CAR expression and inducible cytokines. *Cancers* 12: 375. doi: 10.3390/cancers12020375. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7072617/>

Cancer research and tumor immunology

Engineering of an affinity-matured human antibody that mediates antibody dependent cellular cytotoxicity of mutant p53 expressing cells *in vitro*.

Low, L. *et al.* (2019) Targeting mutant p53-expressing tumours with a T cell receptor–like antibody specific for a wild-type antigen. *Nat. Commun.* 10: 5382. doi: 10.1038/s41467-019-13305-z10. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6879612/>

Antigen-specific CD8⁺ T cell expansion using T Cell TransAct.

Melacarne, A. *et al.* (2021) Identification of a class of non-conventional ER-stress-response-derived immunogenic peptides. *Cell Rep.* 36: 109312. doi: 10.1016/j.celrep.2021.109312. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8278487/>

TCR γ/δ^+ T cells were assessed for their proliferative capacity and cytokine production in the presence or absence of rHMGB1, anti-HMGB1 blocking antibody, acellular MPE fluid, and T Cell TransAct.

Soloff, A. C. *et al.* (2020) HMGB1 promotes myeloid egress and limits lymphatic clearance of malignant pleural effusions. *Front. Immunol.* 11: 2027. doi: 10.3389/fimmu.2020.02027. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7498625/>

T cell immunology

Induction of follicular helper-like (T_{fh}-like) differentiation program in naive human Treg cells *in vitro* using T Cell TransAct, MACS Cytokines, and TexMACS Medium.

Delacher, M. *et al.* (2021) Single-cell chromatin accessibility landscape identifies tissue repair program in human regulatory T cells. *Immunity* 54: 702–720.e17. doi: 10.1016/j.immuni.2021.03.007. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8050210/>

T Cell TransAct reagent used to study Treg cell differentiation and T helper cell subset differentiation under very-low-carbohydrate diet.

Hirschberger, S. *et al.* (2021) Very-low-carbohydrate diet enhances human T cell immunity through immunometabolic reprogramming. *EMBO Mol. Med.* 13: e14323. doi: 10.15252/emmm.202114323. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8350890/>

Co-culture of T Cell TransAct–activated CD4⁺ T cells and CD4⁺CD25⁺CD127^{dim}– Treg cells conditioned with extracellular vesicles (EVs).

Martire, S. *et al.* (2021) A first phenotypic and functional characterization of placental extracellular vesicles from women with multiple sclerosis. *Int. J. Mol. Sci.* 22: 2875. doi: 10.3390/ijms22062875. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC8001892/>

Investigation of previously unrecognized feedback mechanism that regulates T cell activation including expansion of primary T cells using T Cell TransAct.

Yi, J. *et al.* (2019) TCR microclusters form spatially segregated domains and sequentially assemble in calcium-dependent kinetic steps. *Nat. Commun.* 10: 277.
doi: 10.1038/s41467-018-08064-2.
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC6336795/>

Infectious disease research

Development of an antiviral assay system that directly measures anti-HCMV cell-mediated immunity. T Cell TransAct-activated T cells served as positive control.

Houldcroft, C. J. *et al.* (2020) Assessing anti-HCMV cell-mediated immune responses in transplant recipients and healthy controls using a novel functional assay. *Front. Cell. Infect. Microbiol.* 10: 275. doi: 10.3389/fcimb.2020.00275.
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7332694/>

iPSC-derived T cells

Investigation of iPSC-derived T cell functionality (cell proliferation and expression of activation markers) in response to T Cell TransAct-mediated TCR activation.

Netsrithong, R. *et al.* (2020) Multilineage differentiation potential of hematoendothelial progenitors derived from human induced pluripotent stem cells. *Stem Cell Res. Ther.* 11: 481. doi: 10.1186/s13287-020-01997-w.
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC7659123/>



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