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**200 µL CD154 Antibody, anti-human, APC-Vio 770, REAfinity** (clone: REA238, isotype: recombinant human IgG1)

**200 µL IL-2 Antibody, anti-human, PE-Vio 615, REAfinity** (clone: REA689, isotype: recombinant human IgG1)

**2×200 µL CytoStim™, human**

**25 mL Inside Fix**

**2×105 mL Inside Perm**

**2 mL Brefeldin A (100 µg/mL)**

**50 mL Red Blood Cell Lysis Solution (10×)**

**Capacity** 100 tests for up to 100 mL whole blood.

**Product format** Antibodies are supplied in buffer containing stabilizer and 0.05% sodium azide. Brefeldin A is supplied in buffer containing 10% DMSO. Inside Fix contains 3.7% formaldehyde. Inside Perm contains a detergent. CytoStim is supplied in buffer containing stabilizer.

**Storage** Store protected from light at 2–8 °C. Do not freeze. The expiration dates are indicated on the vial labels.

## Warnings

Reagents contain sodium azide. Under acidic conditions sodium azide yields hydrazoic acid, which is extremely toxic. Azide compounds should be diluted with running water before discarding. These precautions are recommended to avoid deposits in plumbing where explosive conditions may develop.

## 1. Description

This product is for research use only.

**Components**

**200 µL CD3 Antibody, anti-human, APC, REAfinity** (clone: REA613, isotype: recombinant human IgG1)

**200 µL CD4 Antibody, anti-human, Vio® Bright B515, REAfinity** (clone: REA623, isotype: recombinant human IgG1)

**200 µL CD8 Antibody, anti-human, VioGreen™, REAfinity** (clone: REA734, isotype: recombinant human IgG1)

**200 µL IFN-γ Antibody, anti-human, PE, REAfinity** (clone: REA600, isotype: recombinant human IgG1)

**200 µL TNF-α Antibody, anti-human, PE-Vio 770, REAfinity** (clone: REA656, isotype: recombinant human IgG1)

**200 µL CD14 Antibody, anti-human, VioBlue®, REAfinity** (clone: REA599, isotype: recombinant human IgG1)

**200 µL CD20 Antibody, anti-human, VioBlue, REAfinity** (clone: REA780, isotype: recombinant human IgG1)

### 1.1 Protocol overview

<b>Stimulation</b>	Transfer 1 mL whole blood sample into 5 mL tubes or well plates. For stimulation add antigen (e.g. PepTivator®), positive control (CytoStim), or negative control. Incubate for at least 8 hours in the presence of Brefeldin A.	8 hours
<b>Red blood cell lysis</b>	Add 1× Red Blood Cell Lysis Solution and incubate for 15 minutes at room temperature (RT) followed by centrifugation and aspiration of supernatant. Wash samples with PEB buffer, centrifuge, and aspirate supernatant.	~ 1.5 hours
<b>Fixation</b>	For fixation add Inside Fix and resuspend. Incubate for 20 minutes at RT.	
<b>Permeabilization and intracellular staining</b>	Centrifuge and aspirate supernatant. Wash with Inside Perm, centrifuge, and aspirate supernatant. Add antibody staining cocktail. Incubate for 10 minutes at RT. Add Inside Perm, centrifuge, and aspirate supernatant. Resuspend in PEB buffer.	
<b>Flow cytometric analysis</b>	Proceed to sample acquisition and analysis.	

## 1.2 Principle

The SARS-CoV-2 T Cell Analysis Kit (Whole Blood), anti-human, REAfinity has been developed for the fast and easy detection of SARS-CoV-2-reactive T cells by intra- and extracellular staining of activation markers and cytokines directly from whole blood samples. The kit contains antibodies for the identification of CD4<sup>+</sup> and CD8<sup>+</sup> T cells, the exclusion of monocytes and B cells as well as the staining of activation markers and cytokines. Furthermore, a positive control (CytoStim), Brefeldin A, Red Blood Cell Lysis Solution for removal of erythrocytes, and reagents for the fixation and permeabilization of cells after stimulation (Inside Fix and Inside Perm) are included. The optimized antibody panel and protocol ensure a comprehensive and efficient analysis of SARS-CoV-2-reactive T cells in minimal amount of whole blood. The kit applies recombinantly engineered REAfinity Antibodies. REAfinity Antibodies are recombinant antibodies that provide superior lot-to-lot consistency and purity compared to mouse or rat hybridoma-derived, monoclonal antibodies. They have been recombinantly engineered to produce highly specific antibodies that require no FcR blocking step. Additionally, they all have the same IgG1 isotype, requiring less isotype controls.

## 1.3 Background information

T lymphocytes execute and control immunological reactions with a repertoire of cytokines, cytotoxic substances, and other mediators. The quantitative and qualitative analysis of CD4<sup>+</sup> T cells and CD8<sup>+</sup> T cells specifically recognizing and reacting towards a defined antigen provides important information to understand their function in various immunological situations. In the context of COVID-19, SARS-CoV-2-reactive T cells are induced. The presence of these cells indicates an infected or convalescent donor and may also allow conclusions on disease progression, severity, specific immune reaction and status. Antigen-reactive T cells can thereby be identified and characterized by analyzing their effector function, e.g., upregulation of activation markers and production of cytokines.

## 1.4 Applications

- Rapid detection of CD4<sup>+</sup> and CD8<sup>+</sup> SARS-CoV-2-reactive T cells from whole blood
- Phenotypical characterization of activated T cells after stimulation with SARS-CoV-2 PepTivator Peptide Pools by flow cytometry
- Immunomonitoring of SARS-CoV-2-reactive T cells
- Research and monitoring of infection- and vaccine-specific T cell responses in individuals

## 1.5 Reagent and instrument requirements

- Phosphate-buffered saline (PBS), pH 7.2, without azide, protein, or other amine-containing compounds.
- PEB buffer: Prepare a solution containing phosphate-buffered saline (PBS), pH 7.2, 0.5% bovine serum albumin (BSA), and 2 mM EDTA by diluting MACS<sup>®</sup> BSA Stock Solution (# 130-091-376) 1:20 with autoMACS<sup>®</sup> Rinsing Solution (# 130-091-222). Keep buffer cold (2–8 °C). Alternatively, use autoMACS Running Buffer (# 130-091-221).

▲ **Note:** EDTA can be replaced by other supplements such as anticoagulant citrate dextrose formula-A (ACD-A) or citrate phosphate dextrose (CPD). BSA can be replaced by other proteins such as human serum albumin, human serum, or fetal bovine serum (FBS). Buffers or media containing Ca<sup>2+</sup> or Mg<sup>2+</sup> are not recommended for use.

- SARS-CoV-2 PepTivators, e.g., Prot\_S (# 130-126-700), Prot\_S1 (# 130-127-041), Prot\_S+ (# 130-127-311), Prot\_N (# 130-126-698), Prot\_M (# 130-126-702). For more information refer to [www.miltenyibiotec.com/peptivators](http://www.miltenyibiotec.com/peptivators).
- Double-distilled water (ddH<sub>2</sub>O).
- Cell culture-grade DMSO.
- 5 mL tubes or a polypropylene deep well plate with round bottom with a minimal volume of 3.5 mL per well.
- Flow cytometer equipped with a red (640 nm), a blue (488 nm), and a violet (405 nm) laser, e.g., MACSQuant<sup>®</sup> Analyzer 10 (# 130-096-343) or MACSQuant Analyzer 16 (# 130-109-803).
- (Optional) MACS Comp Bead Kit, anti-REA (# 130-104-693) for optimal compensation of the fluorescence spillover from fluorochrome-conjugated antibodies.

## 2. Protocol

### 2.1 Reagent preparation

#### Preparation of sterile water/10% DMSO solution

Mix 100 µL cell culture-grade DMSO with 900 µL sterile water to be used to reconstitute the PepTivator as well as for the negative control.

#### Preparation of antibody staining cocktail

For each staining of 1 mL whole blood add 2 µL of each of the nine antibodies to 82 µL of Inside Perm to a total volume of 100 µL per staining.

▲ **Note:** Prepare staining master mix freshly before use. Store staining master mix at 2–8 °C in the dark. Discard unused solution at the end of the day.

#### Reconstitution of PepTivator Peptide Pools

▲ SARS-CoV-2 PepTivator Peptide pools are available in two sizes (6 nmol/peptide and 60 nmol/peptide per vial). The following reconstitution protocol is for the SARS-CoV-2 PepTivator Peptide Pools of the size 6 nmol/peptide.

▲ Please note that 6 nmol/SARS-CoV-2 PepTivator Peptide Pool (1 vial) is sufficient for the stimulation of 10 mL whole blood. The following table shows the amount of PepTivator and quantity of vials needed for different numbers of donors when using 1 mL of whole blood per donor.

**Table 1:** Required amount of PepTivator for different numbers of donors.

	1 donor (1 mL whole blood)	10 donors (10 mL whole blood)	33 donors* (33 mL whole blood)
Amount of PepTivator	0.6 nmol/peptide	6 nmol/peptide	19.8 nmol/peptide
Number of PepTivator vials (1 vial = 6 nmol/ peptide)	1 vial	1 vial	4 vials

\* A maximum of 33 donors can be tested with one kit if each donor is tested for three conditions: antigen stimulation, positive control, and negative control.

- For reconstitution of the lyophilized peptide pool take the vial from –20 °C and warm-up to room temperature.

▲ **Note:** Do not open the vial by removing the rubber plug.

- To dissolve the 6 nmol PepTivator Peptide Pool fill a sterile syringe (0.5 mL) with 500  $\mu$ L of sterile water/10% DMSO solution.
- Slowly inject the sterile water/10% DMSO solution with a sterile needle through the center of the rubber plug into the vial containing the lyophilized peptide pool.
- Vortex the solution to completely dissolve the lyophilized peptide pool. The concentration of the resulting PepTivator stock solution is 12 nmol (approximately 20  $\mu$ g) of each peptide per mL.
- Remove the rubber plug and aspirate the stock solution with a pipette.
- To avoid repeated freeze-thaw cycles prepare working aliquots from the stock solution.
- Store the working aliquots at  $-80^{\circ}\text{C}$ .

#### Preparation 1 $\times$ Red Blood Cell (RBC) Lysis Solution

Dilute Red Blood Cell Lysis Solution (10 $\times$ ) 1:10 with double-distilled water (ddH<sub>2</sub>O). For example, dilute 2 mL of Red Blood Cell Lysis Solution (10 $\times$ ) with 18 mL of ddH<sub>2</sub>O.

▲ **Note:** Do not dilute with deionized water. Store prepared 1 $\times$  Red Blood Cell Lysis Solution at room temperature. Discard unused solution at the end of the day.

#### Preparation of fixation mix

Prepare for each sample of 1 mL whole blood:

- Mix 250  $\mu$ L PEB buffer with 250  $\mu$ L Inside Fix.
- Mix the solution by 3 seconds of vortexing.

▲ **Note:** Prepare fixation mix freshly before use. Store fixation mix at room temperature. Discard unused solution at the end of the day.

#### 2.2 Sample preparation and cell stimulation

▲ For optimal stimulation, use only blood collection tubes containing heparin. EDTA and citrate as anticoagulants impair T cell stimulation.

▲ For optimal results use freshly drawn whole blood stored at room temperature not longer than 24 hours.

▲ Volumes given below are for 1 mL whole blood. Perform three tests for one donor. Use 1 mL for each antigen stimulation, positive control, and negative control.

- Collect at least 3 mL of venous blood in a collection tube containing an appropriate anticoagulant.
- Mix whole blood collection tubes by inverting five times.
- Transfer 1 mL of whole blood into a 5 mL tube or well per sample.
- For antigen stimulation: Add 50  $\mu$ L PepTivator stock solution to the respective 5 mL tubes or wells. Mix by pipetting up and down.

▲ **Note:** The recommended final concentration of PepTivator in the cell suspension is 0.6 nmol (approximately 1  $\mu$ g) of each peptide/mL.

▲ **Note:** When using more than one PepTivator Peptide Pool for stimulation, add equal amounts of each PepTivator to the cell suspension. To avoid further dilution the total maximum amount of added PepTivator should not exceed 100  $\mu$ L per 1 mL whole blood.

- For the positive control: Add 10  $\mu$ L CytoStim, human to the respective 5 mL tubes or wells. Mix by pipetting up and down.

- For the negative control: Add 50  $\mu$ L sterile water/10% DMSO solution to the respective 5 mL tubes or wells. Mix by pipetting up and down.

▲ **Note:** It is recommended to include at least one positive and negative control for each donor.

▲ **Note:** When using more than one PepTivator Peptide Pool for stimulation, adjust volume of the sterile water/10% DMSO solution in the negative control, accordingly.

- Add 20  $\mu$ L of Brefeldin A to each 5 mL tube or well. Mix the samples by pipetting up and down.

▲ **Note:** Thorough mixing is required for a homogeneous stimulation.

- Incubate samples at  $37^{\circ}\text{C}$ , 5% CO<sub>2</sub>, and approx. 95% humidity for 8 hours.

▲ **Note:** Stimulation results may vary between donors and individual PepTivator specificities.

#### 2.3 Staining protocol

▲ Prepare a sufficient amount of 1 $\times$  RBC Lysis Solution, fixation mix, and antibody staining cocktail (refer to chapter 2.1).

▲ If the cell pellet is still red from non-lysed erythrocytes after aspiration of supernatant, the pellet might be “fluffy”. Do not disturb the pellet when aspirating the supernatant.

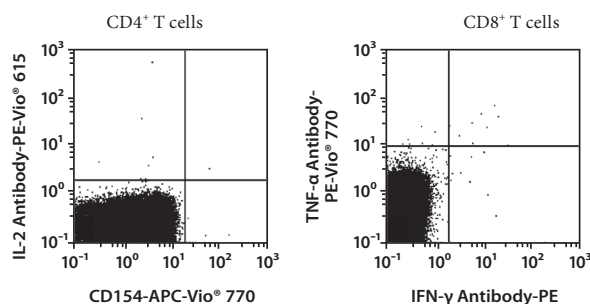
- Remove the samples from the incubator and open the lids.
- Add 2 mL of 1 $\times$  RBC Lysis Solution to each sample.
- Mix by pipetting up and down.
- Incubate for 15 minutes at room temperature (19–25  $^{\circ}\text{C}$ ).
- Centrifuge at 300 $\times$ g for 5 minutes.
- Carefully aspirate supernatant by using a vacuum pump or by pipetting. Do not disturb the pellet.
- Add 1 mL PEB buffer to each sample.
- Mix by pipetting up and down.
- Centrifuge at 300 $\times$ g for 5 minutes.
- Carefully aspirate supernatant by using a vacuum pump or by pipetting.
- Add 500  $\mu$ L fixation mix to each sample.
- Mix by pipetting up and down.
- Incubate for 20 minutes at room temperature (19–25  $^{\circ}\text{C}$ ).
- Centrifuge at 300 $\times$ g for 5 minutes.
- Carefully aspirate supernatant by using a vacuum pump or by pipetting.
- Add 1 mL Inside Perm to each sample.
- Mix by pipetting up and down.
- Centrifuge at 300 $\times$ g for 5 minutes.
- Carefully aspirate supernatant by using a vacuum pump or by pipetting.
- Add 100  $\mu$ L antibody staining cocktail to each sample.
- Mix by pipetting up and down.
- Incubate for 10 minutes in the dark at room temperature (19–25  $^{\circ}\text{C}$ ).
- Add 1 mL Inside Perm to each sample.

24. Mix by pipetting up and down.
25. Centrifuge at 300×g for 5 minutes.
26. Carefully aspirate supernatant completely by using a vacuum pump or by pipetting.  
 ▲ **Note:** The cell pellet should be white and more “solid”. Do not disturb the pellet when aspirating the supernatant.
27. Resuspend the cell pellet in 250 µL PEB buffer.
28. Analyze cell suspension on a suited flow cytometer, e.g., the MACSQuant Analyzer 16 or store cells for up to 24 hours at 2–8 °C in the dark.

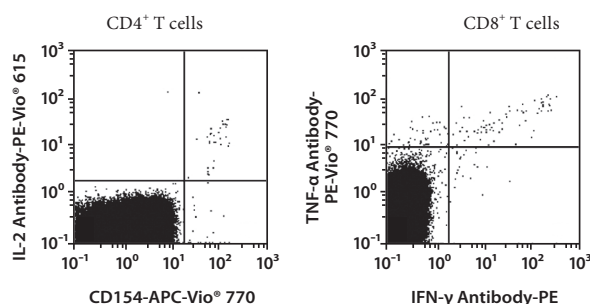
### 3. Example of immunofluorescent staining using the SARS-CoV-2 T Cell Analysis Kit (Whole Blood), anti-human, REAfinity

Human whole blood (1 mL) from SARS-CoV-2-reactive donors were incubated for 8 hours with Brefeldin A. Simultaneously, cells were either left unstimulated (A) or incubated with a selection of SARS-CoV-2 PepTivators Prot\_N (B) or Prot\_S1 (C). Blood samples were lysed, fixed, and permeabilized afterwards. Subsequently, cells were stained with the antibody panel included in this kit. Cells were analyzed using a MACSQuant Analyzer 16. Doublets, debris, and dead cells as well as CD14<sup>+</sup> and CD20<sup>+</sup> cells were excluded. After pre-gating on CD3 as well as CD4 and CD8, respectively, activation marker and cytokine expression were assessed, e.g., CD154 and IL-2 for CD4<sup>+</sup> T cells and TNF-α and IFN-γ for CD8<sup>+</sup> T cells.

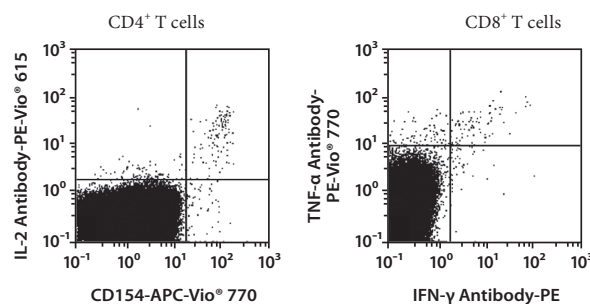
#### A) Unstimulated cells



#### B) Cells stimulated with PepTivator SARS-CoV-2\_Prot\_N



#### C) Cells stimulated with PepTivator SARS-CoV-2\_Prot\_S1



Refer to [www.miltenyibiotec.com](http://www.miltenyibiotec.com) for all data sheets and protocols. Miltenyi Biotec provides technical support worldwide. Visit [www.miltenyibiotec.com](http://www.miltenyibiotec.com) for local Miltenyi Biotec Technical Support contact information.

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