

## CAUTION:

The product as part of the MACS GMP family is for *ex vivo* cell processing only and it is Limited by Federal (or United States) Law to Investigational Use or under an FDA Approval. The product is not intended for human *in vivo* applications.

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## 1. Description

<b>Components</b>	One vial containing 4 mL MACS® GMP T Cell TransAct CR/GMP
<b>Capacity</b>	The reagent is sufficient to activate and expand up to $2 \times 10^8$ enriched T cells or up to $4 \times 10^8$ PBMC in a maximal volume of 70 mL, when used at recommended titer of 1:17.5.
<b>Product format</b>	Polymeric nanomatrix conjugated to recombinant humanized CD3 and CD28 agonist supplied in phosphate buffered-saline (PBS), containing 0.03% poloxamer 188 and 5 g/L recombinant human serum albumin, pH 7.3–7.9
<b>Endotoxin content</b>	$\leq 2$ EU/vial as determined by kinetic Limulus Amebocyte Lysate (LAL) assay (Pharmacopoeia Europaea (Ph. Eur.)).
<b>T cell proliferation assay</b>	$\geq 20\%$ CD25/CD69 co-expression on day 2 $\geq 80\%$ proliferating T cells on day 7
<b>Sterility</b>	Sterility is tested according to Ph. Eur.
<b>Transport</b>	At $+2^\circ\text{C}$ to $+8^\circ\text{C}$ ( $+35^\circ\text{F}$ to $+46^\circ\text{F}$ )
<b>Storage</b>	Store protected from light at $+2^\circ\text{C}$ to $+8^\circ\text{C}$ ( $+35^\circ\text{F}$ to $+46^\circ\text{F}$ ). Do not freeze.
<b>Shelf life</b>	The use-by date is indicated on the vial label.

## Disclaimer

CR/GMP products are for *ex vivo* cell processing only, and are not intended for human *in vivo* applications.

## Quality statement

CR/GMP products are designed, manufactured and tested under an ISO 13485 quality management system and are in compliance with relevant GMP guidelines. They are designed following the recommendations of USP <1043> on ancillary materials. The manufacturing and testing of this product is in compliance with EP chapter 5.2.12 for “Raw materials of biological origin for the production of cell-based and gene therapy medicinal products”.

The declaration of animal- or human-derived materials is given on the Certificate of Origin.

## Background information

Polyclonal T cell expansion can be used when increased numbers of T cells are required or when T cells are activated to enable gene modification. The MACS GMP T Cell TransAct CR/GMP has been designed to activate and expand enriched T cell populations or human resting T cells from peripheral blood mononuclear cells (PBMCs) and is provided in a format directly applicable to CliniMACS® Prodigy.

Due to the nanoscale structure of the MACS GMP T Cell TransAct CR/GMP, it can be sterile filtered and excess reagent is removed by washing. This reagent is suitable for the use in automated culture systems, such as the CliniMACS Prodigy.

## 2. Intended purpose

- The MACS GMP T Cell TransAct CR/GMP is intended for the *in vitro* stimulation and expansion of human T cells from hematological cell populations (e.g. PBMC or CD4 and CD8 enriched T cells).

## 3. Warnings and precautions

- ▲ The instructions for use must be followed.
- ▲ Do not inject or infuse the product directly into humans. Not for human applications.
- ▲ When using this product, the national legislation and regulations must be followed. Any application of *ex vivo* processed target cells is exclusively within the responsibility of the user.
- ▲ For single use only. Do not reuse.
- ▲ Use only if vials are undamaged and sealed.
- ▲ Do not use after the use-by date indicated on the vial label.

## 4. Instructions for use

### 4.1 Reagents and materials required

Product	Order No
MACS GMP T Cell TransAct CR/GMP	200-076-202
TexMACS GMP Medium – 2000mL	170-076-306
MACS GMP Recombinant Human IL-2, or equivalent	170-076-146
MACS GMP Recombinant Human IL-7, or equivalent	170-076-111
MACS GMP Recombinant Human IL-15, or equivalent	170-076-114

### 4.2 General notes

- ▲ Excess MACS GMP T Cell TransAct CR/GMP can be simply removed by a washing step, e.g. medium replacement or centrifugation followed by supernatant replacement (at least 10-fold reduction) 2–3 days after initial stimulation. Performing a washing step earlier may result in reduced T cell proliferation.
- ▲ MACS GMP T Cell TransAct CR/GMP has been developed in combination with TexMACS™ GMP Medium and IL-2 or IL-7 + IL-15.
- ▲ MACS GMP T Cell TransAct CR/GMP can be used in combination with retro- or lentiviral transduction. In case of lentiviral transduction it is recommended to transduce T cells 1 day after stimulation, whereas for retroviral transduction 2–3 days after stimulation.
- ▲ Presence of residual EDTA (e.g. when using CliniMACS buffer) will hamper T cell stimulation. Ensure removal (i.e. over 200-fold reduction) of EDTA prior to T cell stimulation with MACS GMP T Cell TransAct CR/GMP e.g. via a washing step.

### 4.3 Protocol

#### 4.3.1 Fresh cell product

For stimulation of fresh PBMCs or enriched T cells a final dilution of 1:17.5 is recommended. For optimal stimulation with the CliniMACS Prodigy T cells should be activated with  $1 \times 10^6$  T cells per  $\text{cm}^2$  respectively  $1.43 \times 10^6$  T cells per mL. For PBMC optimal stimulation conditions are  $2 \times 10^6$  PBMC per  $\text{cm}^2$  respectively  $2.86 \times 10^6$  PBMC per mL. Minimal and maximal cell densities for the stimulation of T cells or PBMC are given below. It is recommended to use the same dilution of 1:17.5 for each condition.















Recommended cell densities for cell activation		
purified T cells	per $\text{cm}^2$	per mL*)
minimum	$0.2 \times 10^6$	$0.28 \times 10^6$
optimum	$1 \times 10^6$	$1.43 \times 10^6$
maximum	$2 \times 10^6$	$2.86 \times 10^6$
PBMC	per $\text{cm}^2$	per mL*)
minimum	$0.4 \times 10^6$	$0.56 \times 10^6$
optimum	$2 \times 10^6$	$2.86 \times 10^6$
maximum	$4 \times 10^6$	$5.72 \times 10^6$

\*) for initial stimulation in the CliniMAC Prodigy TCT application a total of 70 mL culture medium is used

#### 4.3.2 Frozen and thawed cell product

When working with frozen material, it is recommended to rapidly thaw the cells, wash away the freezing solution containing DMSO, resuspend the cells in culture medium at densities below  $5 \times 10^6$  cells / mL and rest the cells overnight at  $+37^\circ\text{C}$  ( $+99^\circ\text{F}$ ) and 5%  $\text{CO}_2$  in culture medium. After recovery, harvest, count, resuspend and stimulate the cells as described in 4.3.1.

## 5. Glossary of symbols

	Manufacturer		Use-by date
	Order number		Do not reuse
	Part number		Temperature limit
	Batch code		Phone
	Consult instruction for use		Fax
	Do not use if package is damaged.		E-mail
	Caution		Website

This data sheet and corresponding information as well as special protocols can be found under [www.miltenyibiotec.com/200-076-202](http://www.miltenyibiotec.com/200-076-202).

### Warranty

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